

AD _____

Award Number: W81XWH-09-1-0432

TITLE: Molecular Regulation of Endothelial Cells by NF-1

PRINCIPAL INVESTIGATOR: Kevin Pumiglia

CONTRACTING ORGANIZATION: Albany Medical College
Albany, NY 12208

REPORT DATE: 2009-06-01

TYPE OF REPORT: Other

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

| | | | | | |
|--|------------------|--|--------------------------------------|--|--|
| REPORT DOCUMENTATION PAGE | | | Form Approved OMB No. 0704-0188 | | |
| Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS. | | | | | |
| 1. REPORT DATE Raj 10 1H | | 2. REPORT TYPE Qj a | | 3. DATES COVERED 15 JUN 00/JAFI 00^8\ à^!00FG | |
| 4. TITLE AND SUBTITLE Molecular Regulation of Endothelial Cells by NF-1 | | 5a. CONTRACT NUMBER | | | |
| | | 5b. GRANT NUMBER W81XWH-09-1-0432 | | | |
| | | 5c. PROGRAM ELEMENT NUMBER | | | |
| 6. AUTHOR(S) Kevin PumigliaZU00E E-Mail: rwo k mB o ckto e0f w | | 5d. PROJECT NUMBER | | | |
| | | 5e. TASK NUMBER | | | |
| | | 5f. WORK UNIT NUMBER | | | |
| 7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Albany Medical College Albany, NY 12208 | | 8. PERFORMING ORGANIZATION REPORT NUMBER | | | |
| 9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012 | | 10. SPONSOR/MONITOR'S ACRONYM(S) | | | |
| | | 11. SPONSOR/MONITOR'S REPORT NUMBER(S) | | | |
| 12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited | | | | | |
| 13. SUPPLEMENTARY NOTES | | | | | |
| 14. ABSTRACT Y ^A00^A ç•d æ^a0A [\& æA^*~ æj } A A0FA A@ { æA } a[@ æA^ •EAV a *A a æ^& A [a^!A ^A00^A^) ^!æ^aa a^ a^ &a ^A } [& a[, } A A0FA a &0æac^!a^a0A^~&0A } & æA a } æj *E æ^!æj } E a aæ & æA [!] @ *^) ^•a EA V@•^Aæa00^A^) A^ a a @ a^A a a } ^) a^aA æ^ •&a EA ^A00^A^c! { a^aA00^A VUUA a } æ^!æj •a^ &a } A A00^Aæ0A & [] [] ^) A A0A0FA^• [] •^A a A00^A ç•d æ^a0A [\A^A VUUA A^*~ æj *A) a[@ æA^ A [!] æ^!æj } A aA { [!] @ *^) ^•a EA ^A00^A^•aA [cA aU00A aA aU00A] ! æ@•A ç•d æ^a0A [\A^A aA [!] { •A A0A0FA { ^aæ^aA^~&0EA ^A00^A^*^•A00^A @ A0E aA0A a &a ^A [!] æ^!æj } A A00^A aA } A^•A00^A A A0A0FA a [!] æ^!æj } A aA a^!aæ & æA [!] @ *^) ^•a EA c^A ^A00^A^ &0^a^aA A^) ^!æj *A a^•^ } æA [^•^A [a^!A00^A [, •A^~&a^ a^ &a ^A^ { [çA^A A0FA A0Aæ & æA } a[@ a { EA^!a a æ^A c^!a^) •A aæaA00^A [••A A0FA A0A } a[@ a { A æA^A^~æ } A A [!] ç [^A } a[@ æA^ A [!] æ^!æj } A A c a *Aæ & æA^•EA0A @ A^~ A [a^!A A0FAæ & æA aæ^A { [çA *A , æaE | | | | | |
| 15. SUBJECT TERMS Endothelial, NF-1, Ras, vasculopathy, molecular, signaling | | | | | |
| 16. SECURITY CLASSIFICATION OF: | | | 17. LIMITATION OF ABSTRACT UU | 18. NUMBER OF PAGES A-I | 19a. NAME OF RESPONSIBLE PERSON USAMRMC |
| a. REPORT U | b. ABSTRACT U | c. THIS PAGE U | | | 19b. TELEPHONE NUMBER (include area code) |

Table of Contents

| | <u>Page</u> |
|---------------------------------------|-------------|
| Introduction..... | 1-2 |
| Body..... | 3-7 |
| Key Research Accomplishments..... | 7 |
| Reportable Outcomes..... | 8 |
| Conclusion..... | 9 |
| References..... | 10-11 |
| Appendices.....(supporting data)..... | 12- 23 |
| (Manuscript)..... | 24-33 |

Introduction:

NF1 and Vascular Disease

Mutations in the *NF1* gene cause Neurofibromatosis type 1, an autosomal dominant disease that affects approximately 1 in 3000 individuals, making it one of the most common inherited genetic disorders. The autosomal dominant genetic inheritance pattern, as well as the mutational predisposition of this locus makes it highly likely the prevalence of this disease will continue to increase in the population (McClatchey 2007). NF1 has variable clinical manifestations. Most commonly changes in skin pigmentation (café au-lait spots as well as freckling) and the presence of benign and malignant nerve sheath tumors termed neurofibromas, are observed (McClatchey 2007). Importantly, a significant clinical manifestation of NF1 disease can include vascular disease. Patients with NF1 disease make up a significant portion of all those patients presenting with renal artery stenosis and early-onset cerebral vascular disease (Friedman, Arbiser et al. 2002). Moreover, cardiovascular disease is a significant contributor to premature death in NF1 patients and is a particular risk factor among younger patients. One recent study suggested that occurrence of vasculopathy was over seven times more likely to occur in NF1 patients under 30 compared to their unaffected peers. (Rasmussen, Yang et al. 2001)

The manifestation of vasculopathy in NF1 affected individuals can be quite variable. Perhaps the most common is renal artery stenosis and hypertension. The renal artery is also a site of aneurysms and aortic coarctation (Zochodne 1984). Recently a model was developed in the Epstein lab where NF1 was knocked out specifically in the smooth muscle cells of mice using the conditional excision of a floxed allele. The basal morphology and structure of these blood vessels appeared to be normal in these mice. However, in response to injury, these mice had exaggerated responses to vessel injury characterized by smooth muscle cell proliferation and vascular stenosis. These changes were accompanied by activation of Ras signaling, thus Ras driven smooth hyperproliferation is thought to be an important contribution to large vessel stenosis phenotypes (Xu, Ismat et al. 2007).

Another site of frequent vascular involvement is the cerebral circulation. These lesions are often occlusive in nature, perhaps as a result of a pro-thrombotic vasculature. They can also involve nodules of mesodermal cells that occlude the vessel and/or distort the vessel wall. Moyamoya-type lesions with accompanying telangiaectasia are sometimes seen, as are arteriovenous fistulae and aneurysms (Friedman, Arbiser et al. 2002). A frequent and debilitating consequence of many of these changes is vascular hemorrhage. Intracranial hemorrhage is associated with headaches, seizures, loss of motor control, and other neurological deficits. It is important to point out that intracerebral micro-hemorrhage has also been associated with loss of cognitive function (Schneider 2007) and could plausibly be involved with the learning deficits often observed in NF1 patients, though this remains unstudied. The risk of a compromised vasculature is exacerbated by any underlying hypertension, as hypertension itself is a risk factor for cerebral hemorrhage. Indeed hemorrhage is the cause of death in upwards of 50% of all patients who die with NF1 associated cerebrovascular disease (Friedman, Arbiser et al. 2002). Thus, a better understanding of the role of NF1 in the vasculature is

an essential requirement in designing better tools to diagnose this disease and reduce its morbidity.

Our Hypothesis and Previous work:

As noted above, numerous pieces of evidence suggest that NF1 plays an important role in regulating the vascular endothelium. NF1 is clinically associated with multiple vasculopathies including malformations, aneurysms, hypertension and consequently there is a markedly elevated risk of cerebrovascular accidents (Friedman, Arbiser et al. 2002). Mouse models have shown that endothelial-selective ablation of NF1 results in embryonic lethality (Gitler, Zhu et al. 2003), which can be rescued by expression of the GRD of NF1 (Ismat, Xu et al. 2006), suggesting Ras activation is critical. Our published data have determined that Ras is required for endothelial responses to VEGF (Meadows, Bryant et al. 2001) and that overexpression of an activated allele of Ras is sufficient to drive multiple angiogenic phenotypes (Meadows, Bryant et al. 2004). We have also observed chronic Ras activation induces abnormal vascular morphogenesis (Bajaj, Zheng et al. 2010). Consistent with this, NF1 haploinsufficient mice show exaggerated angiogenic responses (Wu, Wallace et al. 2006) and data have been published suggesting that shRNA mediated knockdown of NF1 can result in exaggerated activation of signaling in response to growth factors in human endothelial cells (Munchhof, Li et al. 2006). *Our central hypothesis is that NF1 mediated activation of Ras is a critical mediator of vasculopathy associated with NF1 disease.* Our data to date demonstrates that Ras activation phenocopies NF1 deficiency in cell culture models. Furthermore, we find that inducible expression of activated Ras in the adult mouse results in intracranial hemorrhage. However, to prior to this project, specific details on the role of Ras, Ras isoforms and NF1 were unknown. Similarly, there was little to no data on the sufficiency of NF1 loss to alter endothelial cell function. In this project we investigated the molecular regulation of endothelial cell function by NF1 with two Specific Aims: *1) Investigate the relationship between NF1 and Ras isoforms in primary human endothelial cells in vitro; and 2) Determine the consequences of NF1 loss on the vascular endothelium in vivo.*

Key research accomplishments in the first two years include:

- Creating human endothelial cells with drug inducible knockdown of NF1
- Characterization of signal transduction following knockdown of NF1
- Consequences on endothelial cell functions following knockdown of NF1
- Linkage of NF1 loss to activation of mTOR signaling and a role for this signal in the functional outcomes of NF1 loss
- Validation of isoform specific antibodies
- Identification of siRNA and shRNA sequences targeting individual Ras isoforms
- Establishment of NF1^{Flox} and VE: Cad cre mouse colonies
- Establishment of histological staining techniques

Below is a summary of our significant findings, progress, and problems encountered in Year 3 of support as well as a 6 month extension period. This last period was a no-cost extension where we were principally trying finish some outstanding experiments in the

statement of work as well as begin at least a preliminary characterization of our progress on a mouse model.

Body:

A major objective of our last funding year, as outlined in our Statement of Work, is the finalization of our experiments investigating the cellular consequences of NF1 loss in endothelial cells. We have continued building on previous data demonstrating effective NF1 knockdown in endothelial cells with multiple independent constructs. This data was published. The abstract of this work is as follows:

Neurofibromatosis is a well-known familial tumor syndrome, however these patients also suffer from a number of vascular anomalies. The loss of NF1 from the endothelium is embryonically lethal in mouse developmental models, however little is known regarding the molecular regulation by NF1 in endothelium. We investigated the consequences of losing NF1 expression on the function of endothelial cells using shRNA. The loss of NF1 was sufficient to elevate levels of active Ras under nonstimulated conditions. These elevations in Ras activity were associated with activation of downstream signaling including activation of ERK, AKT and mTOR. Cells knocked down in NF1 expression exhibited no cellular senescence. Rather, they demonstrated augmented proliferation and autonomous entry into the cell cycle. These proliferative changes were accompanied by enhanced expression of cyclin D, phosphorylation of p27KIP, and decreases in total p27KIP levels, even under growth factor free conditions. In addition, NF1-deficient cells failed to undergo normal branching morphogenesis in a co-culture assay, instead forming planar islands with few tubules and branches. We find the changes induced by the loss of NF1 could be mitigated by co-expression of the GAP-related domain of NF1, suggesting Ras regulation was responsible for these changes. Inducible knockdown of NF1 shows that the morphogenic changes are reversible upon removal of the shRNA induction. Similarly, in fully differentiated and stable vascular-like structures, the silencing of NF1 results in the appearance of abnormal vascular structures. Finally, the proliferative changes and the abnormal vascular morphogenesis are normalized by low-dose Rapamycin treatment. These data provide a detailed analysis of the molecular and functional consequences of NF1 loss in human endothelial cells. These insights may provide new approaches to therapeutically addressing vascular abnormalities in these patients while underscoring a critical role for normal Ras regulation in maintaining the health and function of the vasculature.

This manuscript was published in PLOS-One in its final form November 7, 2012. The decision to publish there was based on our belief that publicly funded research should be publically available without restriction. This article is freely available at <http://www.plosone.org/article/info%3Adoi%2F10.1371%2Fjournal.pone.0049222> and a copy of it is also appended to this report beginning on page 23.

In the course of these studies we conducted many experiments in control cells to understand the mTOR interventions we were employing to inhibit mTOR in the NF1 cells. This generated novel results and findings that we have split off into a separate manuscript that we are preparing to submit. The principal author of this manuscript abruptly left the lab and graduate program in early October, just as this manuscript was

being about to be submitted. We found inadequate documentation of some of the experiments presented in this manuscript in the available research record from this student. Therefore while this manuscript is written and ready for submission, we have held up submission of this manuscript while a technician in the lab independently replicates the results to make sure the data are completely accurate prior to publication (this replication is being financially supported by the institution). The abstract for that paper reads as follows:

Angiogenesis is important for embryogenesis, wound healing and is a critical element in cancer progression. The mammalian target of rapamycin (mTOR) is a serine threonine kinase, that exists in two distinct signaling complexes: mTORC1 and mTORC2. Aberrant mTOR signaling is often associated with cancer pathologies including renal cell carcinoma, tuberous sclerosis, and neurofibromatosis, which are known to have associated hypervascularity and other dysfunctional vasculature. The regulation of this axis is also disturbed by metabolic abnormalities such as diabetes, with known vascular complications. Thus mTOR may be a significant regulator of endothelial cell functions, including angiogenesis. Given the connections between angiogenesis, vascular dysfunction, and mTOR signaling, we investigated the role of mTORC1 in vascular endothelial cells. We identified a dose of rapamycin (an allosteric inhibitor of mTORC1) for selective inhibition of mTORC1/S6 without disrupting mTORC2/Akt signaling. In addition, we have developed inducible shRNA to knockdown the requisite TORC1 component, Raptor. We find that mTORC1 inhibition affects developing vessels but not established vessels. These effects are largely due to an mTORC1-dependent contribution to cellular proliferation. Our data suggest a role for mTORC1 in the phosphorylation and destabilization of the cyclin-dependent kinase inhibitor, p27^{Kip}. We find identical results using both methods to interfere with TORC1. These data suggest that drugs acting specifically on mTORC1 may provide selective anti-angiogenic activity on developing vasculature through modulation of endothelial cell proliferation, with minimal direct effect on cell survival. These results are of value in developing strategic therapeutic interventions in inherited and pathological conditions where mTOR signaling is enhanced in endothelial cells and contributes to vascular dysfunction.

The figures for this paper are included in this report and comprise *Figures 1-8*, found on pages 12-13.

Another major objective in Aim One of the proposal was to investigate the role of individual Ras isoforms in the endothelial cells, particularly with respect to the loss of NF1. We have successfully knocked down specific isoforms of Ras in primary endothelial cells and as shown in *Figure 9*, we find that the N-Ras isoform is the primary isoform regulated by NF1, while knockdown of the H or K-Ras has little effect of Ras-GTP levels in NF1 knockdown cells. This finding was confirmed when we performed double knockdown experiments such as those shown in *Figure 10*. Similarly, we find that signal transduction that is elevated following the loss of NF1, is only significantly affected by the loss of N-Ras, as shown in *Figure 11*, and in agreement with the GTP loading experiments. To investigate the Ras isoforms in longer term experiments, and to confirm our findings with siRNA, we also developed lentiviral vectors to knockdown Ras isoforms with shRNA in a stable fashion. These constructs were engineered to be able to induce knockdown of Ras isoforms in an inducible fashion under the control of

doxycycline. An example of the knockdown of these isoforms is shown in *Figure 12*, alone and in combination. When we performed experiments with these vectors in normal endothelial cells, the formation of vascular structures did not rely on N-Ras or H-Ras, but rather was dependent upon K-Ras, as shown in *Figure 13*. In the absence of K-Ras, cells did not appear to elongate and anastomose properly. In agreement with these findings we found that even in the presence of double knockdown of H and N-Ras, normal vascular structures could form (*Figure 14*). As NF1 activated N-Ras preferentially, we also wanted to test the consequences of N-Ras activation compared to other isoforms. We used lentiviral infection with viruses that expressed activated mutants of Ras under inducible control as shown in *Figure 15*. Endothelial cells, infected with these constructs all made normal tubes under non-induced conditions (-Dox), however upon expression of the active mutant, changes in vascular morphogenesis were noted, *Figure 16*. Interestingly activation of H-Ras had the most profound phenotype, completely abrogating vascular morphogenesis and promoting the formation of sheet-like formations. In contrast K-Ras, which is required for tube formation as shown in *Figures 13 and 14*, had no significant effect on the morphology of tubes when constitutively activated. Consistent with our results with NF1, activation of N-Ras gave an intermediate phenotype with significant defects in vascular morphogenesis, but with some capacity to still sprout tubes. We next wanted to probe the role of the Ras isoforms in the generation of the phenotype seen with NF1 loss. Here we generated cells that were double infected with a vector that knocks down NF1 (co-expresses GFP) and one that knocks down the desired Ras isoform upon induction with doxycycline (RFP). Using dual color sorting we isolated cells that were doubly positive and analyzed their phenotypes. Here we obtained surprising and at this point perplexing results. As noted in *Figure 17*, when we silenced N-Ras in NF1 expressing cells, there was little restoration of proliferation to baseline levels as would have been predicted, as NF1 loss seemed to activate N-Ras. In contrast, while NF1 showed no activation of H-Ras, loss of H-Ras restores normal baseline proliferation. Interestingly, we found a similar pattern in the phosphorylation of p27 as shown in *Figure 18*, a molecule which we believe plays an important role in the proliferative response to NF1 based on our published data (Bajaj, Li et al. 2012). Finally we investigated these cells in the vascular morphogenesis assay. While the double infected with NF1 knockdown and a non-targeting control show a greatly diminished capacity to form tubes and instead form sheet like-structures, silencing of N-Ras did little to improve the tube forming ability of these cells. In contrast, silencing H-Ras in the NF1 knockdown cells provides a robust recovery of the tube forming capability of these cells *Figure 19*. Thus these data are consistent across all three assays, showing an important role for H-Ras and K-Ras. However these data appear at the surface inconsistent with the data obtained using si-RNA based targeting of the Ras isoforms, where H-Ras was not activated at all, and N-Ras demonstrated the strongest level of activation. There are numerous technical and mechanistic possibilities for these findings that we are currently trying to work through. We need to insure that the Ras activation profile is the same in the shRNA silenced cells, as they are silenced for a long time and it may be that some isoforms may be more able to be compensated for. For example, silencing of N-Ras may result in a stronger activation of H and K-Ras that is acquired over time in cells with low or no NF1. In contrast, silencing H-Ras may not result in any compensatory activation of N-Ras. Another possibility is that the most important Ras

family GTPase is not one of the major isoforms but rather a little known Ras family member such as TC-21. Indeed this protein was recently shown to be activated in NF1-deficient glial cells (Patmore, Welch et al. 2012). While these unexpected results slowed submission of a third manuscript (as it is no longer a nice neat N-Ras related story) we are confident in the months ahead we will better understand the mechanistic basis for these findings and as such be in a better position to publish them with meaningful and correct interpretation.

As noted in our original statement of work, we also were developing an in vivo model of NF1 loss in the endothelium. We have had significant delays and problems with this, largely as a consequence of a published strain of mice (Monvoisin, Alva et al. 2006) that we proposed to use originally to provide the endothelial specific excision of NF1 not performing in adult mice consistent with the published data (performed in neonates). After trying different induction protocols, we bred in an integrated reporter strain (Rosa26-td-Tomato), in the hope that we could track affected cells. However a preliminary experiment revealed no obvious phenotype and only sparsely detectable cells. We realized this was not going to allow an efficient evaluation either. Fortunately, in the last year we have been successful in importing and establishing a colony of another recently available strain that is inducible, has an endothelial-specific Cre. We have bred that strain onto the integrated Cre-reporter strain, ROSA26-LSL-Td-Tomato, which upon successful recombination expresses the red fluorescent protein td-Tomato. As shown in *Figure 20*, upon tamoxifen treatment we can demonstrate highly penetrant expression (approximately 80-85%) of td-tomato that co-localizes with endothelium as indicated by CD31 staining. Thus, for the first time we have a model where we can induce recombination of a floxed gene with high efficiency in the adult mouse (embryonic recombination of NF1 is known to be lethal). As such we have begun the cross breeding to the NF1^{flox} and have generated mice with all three desired alleles, as shown in *Figure 21*. We have conducted a basic characterization of a limited number of NF1^{flox/flox} mouse by the termination of the grant term. In these mice, we have treated littermates with vehicle or Tamoxifen to induce the endothelial-specific Cre. After 30 days the mice were sacrificed. We have performed some preliminary analysis of the blood vessels in these mice in retinal whole mounts, as these are an excellent vascular bed to visualize vascular patterning. We found excellent recombination in the tri-genic strain, similar to the bi-transgenic we analyzed in *Figure 20*. To date we have not observed any gross vascular abnormalities in patterning (aneurysm, malformations, and dilations) or hemorrhage in the 30 day period. However we have detected what appears to be an increase in endothelial cell proliferation as shown by Ki-67 staining, *Figure 22*. It is important to keep in mind that in this model both genes need to be floxed for the cell to lose NF1 expression and while the penetrance of the Cre looks good, it is impossible to determine visually the extent of NF1 excision in the endothelium. While this project is over we are pleased that the data we were able to obtain developing this model was useful in generating a successful application to the CDMRP-NF1 program in which work on a vascular model of NF1 loss will be continued. In these future experiments we will extend this model by breeding it to the NF1 deletion strain which lacks one copy of the NF1 gene from birth. This should genocopy the human lesions, and increase the frequency of complete loss of NF1 in the endothelium compared to the flox/flox model. In addition, it

will replicate the haplo-insufficiency of the stromal cells which may contribute to the pathogenesis, as endothelial cells are very responsive to micro-environmental changes (Le and Parada 2007, Reilly and Van Dyke 2008, Kumar and Weaver 2009).

We are optimistic that these future experiments will be successful as we are well prepared to characterize the vascular phenotype of the mouse. We have worked out all of the histological staining to detect Ki-67, CD31, phospho-ERK, Phospho-S6, inflammatory cells, red blood cells, basement membrane, and pericytes. We have learned how to prepare retinal whole mounts, a technique that is very sensitive for the detecting abnormalities in vascular patterning. In addition, we learned how to isolate purified endothelial cells from the lung and culture them, as well as generate microvessel fragments from the brain, where we anticipate a phenotype may reveal itself. These microvessels, shown in *Figure 23*, allow biochemical characterization of a vascular enriched component following rapid isolation. We have verified that these can be used for the detection of active Ras and Ras related signals using biochemical approaches (as shown in *Figures 24 and 25*.) These will all be necessary analysis in evaluating the effects of NF1 loss in the vasculature of the mouse model moving forward.

Key Accomplishments in Year 3 and extension:

- Completion a manuscript detailing analysis of cellular signaling and function in response to NF1
- Completion of analysis and preparation of a manuscript determine the role of mTOR in endothelial cell function
- Finalization of studies using shRNA that implicate all three isoforms in apparently distinct ways in the process of normal vascular morphogenesis.
- Determination that N-Ras hyperactivation is sufficient to induce abnormal vascular morphogenesis
- Performance of experiments investigating the loss of individual Ras isoforms on the phenotype of NF1 deficient cells. These experiments conducted in dual knockdown primary cell cultures revealed an unexpected important role for H-Ras in the proliferative and tubulogenic defects observed following loss of NF1 expression
- Final determination to abandon the breeding of the *VE-Cad:CRE-ERT²* strain due to poor/hard to document recombination
- Importation of mouse, colony establishment and demonstration that CAD5-CRE^{ERT2} gives high efficiency recombination
- Preliminary analysis of *CAD5-Cre^{ERT2}/Rosa26-LSL-td-Tomato/NF1^{flox/flox}* mice and demonstration that after 30 days these mice have regions of enhanced endothelial cell proliferation (Ki-67 staining).
- Establishment of techniques such as cell purification and micro-vessel fragment isolation to permit ex vivo analysis of biochemical properties of NF1^{flox/flox} endothelium. Particularly useful in future studies
- Successful competition for CDMRP-NF1 program support for continued investigation into the mechanistic basis of vascular dysfunction in the endothelium of NF1 patients, including support for continued generation of a mouse model.

Reportable Outcomes:

Publications:

Loss of NF1 Expression in Human Endothelial Cells Promotes Autonomous Proliferation and Altered Vascular Morphogenesis Bajaj, A., Li Q-F, Zheng, Q. and Pumiglia, K. [PLOS One](#)

TORC1 has a Critical Role in Endothelial Cell Proliferation and Morphogenesis
Zheng, Q. and Pumiglia, K [PLOS One](#) (to be submitted spring 2013)

Ras isoforms are distinctly regulated by NF1 and play unique roles in the abnormal Vascular morphogenesis following the loss of NF1. Li, Q-F., Zheng, Q., and Pumiglia K. (Manuscript in preparation).

Personnel Supported:

Kevin Pumiglia Ph.D. - Principal Investigator

Anshika Bajaj Ph.D. –Postdoctoral fellow (left in summer 2010 for staff position at GE Life Sciences)

Qing-Fen Li Ph.D. - Post-doctoral Fellow

Qingxia Zheng M.S. - Research Technician

Shawn Hakim M.S. - Graduate student (withdrew 10/2012, prior to degree conferral)

Unique Reagents Generated:

- Lentivirus vectors to express Mir-based shRNAs targeting NF1 expression, under doxycycline inducible control
- Lentivirus vectors to express Mir-based shRNAs specifically targeting H, K, and N-Ras expression, under doxycycline inducible control
- Lentivirus vectors to express activated mutants of each Ras isoform
- Lentivirus vector to express the GAP-domain of NF1
- Lentivirus expressing Mir-based shRNAs targeting RAPTOR, an essential protein in the mTOR pathway
- Unique strains of mice containing Endothelial specific Cre recombinase under tamoxifen inducible control with integrated reporter gene (ROSA26-LSL:td-Tomato) as well as this mice with the NF1 gene replaced by a floxed copy

Funding Applied For:

NIH – *NF1 and endothelial dysfunction* (2012): Not funded

DOD-CDMRP- *NF1 Signal Transduction and Vascular Dysfunction* (2012): NF120051, recommended for funding. Start date pending.

Conclusions:

We have completed essentially all of the experiments originally proposed in the “*Statement of Work*”. We continue to replicate these data in a few instances to prepare these data for publication or obtain sufficient representative data for quantification. One manuscript has been published, another is completely written and merely awaiting some confirmatory experimentation. A third has a large body of data associated with it but due to some of the unanticipated results some additional experimentation will have to be done to provide an accurate interpretational context. We believe these are the most comprehensive and molecularly sophisticated studies to date on the role of NF1 in endothelial cells and perhaps in any cell. Our studies provide a broad view of the range of endothelial cell functions impacted by NF1 knockdown. They directly implicate N-Ras as a critical player in these abnormal signaling events but also revealed a surprising role for H-Ras given its apparent lack of activation following loss of NF1. Our data directly implicate mTORC1 as a critical molecular signal in the pro-growth and vascular dysmorphogenic phenotypes. This information may be highly critical as there are numerous FDA approved therapeutics available to target this pathway. A more comprehensive understanding of the relationship of mTOR signaling to vascular endothelial dysfunction in NF1 patients may lead to novel ways to intervene medically in NF1 associated vasculopathies. The isoform studies will also prove beneficial to NF1 patients, as Ras isoforms are differentially affected by several classes of drugs including the widely prescribed “statins”, as well as several experimental compounds. A better understanding of the relationship between the Ras isoforms and dysfunction mediated by loss of NF1, will provide guidance on whether some of these regimens might be beneficial (or harmful) to NF1 patients. Lastly, the ultimate test of many of our hypotheses regarding vascular dysfunction in NF1 patients will be in the complex tissue microenvironment. This requires *in vivo* experimentation. We have generated animals capable of high efficiency recombination in the endothelium. We have detected an increase in the proliferation of endothelial cells in the vasculature of mice induced to remove the NF1 allele suggesting that indeed the loss of NF1 in the endothelium can lead to vascular defects. While these studies have progressed slower than we wished, they will provide the first animal model of NF1 loss in the mature vascular endothelium. This animal model will lead to novel insights into this important disease. We have also accumulated the requisite skills for the IHC and biochemical characterization of these mice in the coming years. We look forward to revealing the role of NF1 in the vascular endothelium in the coming years as a consequence of our successful competition for funding to continue to investigate this important aspect of biology. These studies are critical for the best management of NF1 patients.

References:

- Bajaj, A., Q. F. Li, Q. Zheng and K. Pumiglia (2012). "Loss of NF1 expression in human endothelial cells promotes autonomous proliferation and altered vascular morphogenesis." PLoS One **7**(11): e49222.
- Bajaj, A., Q. Zheng, A. Adam, P. Vincent and K. Pumiglia (2010). "Activation of endothelial ras signaling bypasses senescence and causes abnormal vascular morphogenesis." Cancer Res **70**(9): 3803-3812.
- Friedman, J. M., J. Arbiser, J. A. Epstein, D. H. Gutmann, S. J. Huot, A. E. Lin, B. McManus and B. R. Korf (2002). "Cardiovascular disease in neurofibromatosis 1: report of the NF1 Cardiovascular Task Force." Genet Med **4**(3): 105-111.
- Gitler, A. D., Y. Zhu, F. A. Ismat, M. M. Lu, Y. Yamauchi, L. F. Parada and J. A. Epstein (2003). "Nf1 has an essential role in endothelial cells." Nat Genet **33**(1): 75-79.
- Ismat, F. A., J. Xu, M. M. Lu and J. A. Epstein (2006). "The neurofibromin GAP-related domain rescues endothelial but not neural crest development in Nf1 mice." J Clin Invest **116**(9): 2378-2384.
- Kumar, S. and V. Weaver (2009). "Mechanics, malignancy, and metastasis: The force journey of a tumor cell." Cancer and Metastasis Reviews **28**(1-2): 113-127.
- Le, L. Q. and L. F. Parada (2007). "Tumor microenvironment and neurofibromatosis type I: Connecting the GAPs." Oncogene **26**(32): 4609-4616.
- McClatchey, A. I. (2007). Neurofibromatosis. Annual Review of Pathology. **2**: 191-216.
- Meadows, K. N., P. Bryant and K. Pumiglia (2001). "Vascular endothelial growth factor induction of the angiogenic phenotype requires Ras activation." J Biol Chem **276**(52): 49289-49298.
- Meadows, K. N., P. Bryant, P. A. Vincent and K. M. Pumiglia (2004). "Activated Ras induces a proangiogenic phenotype in primary endothelial cells." Oncogene **23**(1): 192-200.
- Monvoisin, A., J. A. Alva, J. J. Hofmann, A. C. Zovein, T. F. Lane and M. L. Iruela-Arispe (2006). "VE-cadherin-CreERT2 transgenic mouse: a model for inducible recombination in the endothelium." Dev Dyn **235**(12): 3413-3422.
- Munchhof, A. M., F. Li, H. A. White, L. E. Mead, T. R. Krier, A. Fenoglio, X. Li, J. Yuan, F. C. Yang and D. A. Ingram (2006). "Neurofibroma-associated growth factors activate a distinct signaling network to alter the function of neurofibromin-deficient endothelial cells." Hum Mol Genet **15**(11): 1858-1869.
- Patmore, D. M., S. Welch, P. C. Fulkerson, J. Wu, K. Choi, D. Eaves, J. J. Kordich, M. H. Collins, T. P. Cripe and N. Ratner (2012). "In vivo regulation of TGF-beta by R-Ras2 revealed through loss of the RasGAP protein NF1." Cancer Res **72**(20): 5317-5327.
- Rasmussen, S. A., Q. Yang and J. M. Friedman (2001). "Mortality in neurofibromatosis 1: an analysis using U.S. death certificates." Am J Hum Genet **68**(5): 1110-1118.
- Reilly, K. M. and T. Van Dyke (2008). "It Takes a (Dysfunctional) Village to Raise a Tumor." Cell **135**(3): 408-410.
- Schneider, J. A. (2007). "Brain microbleeds and cognitive function." Stroke **38**(6): 1730-1731.
- Wu, M., M. R. Wallace and D. Muir (2006). "Nf1 haploinsufficiency augments angiogenesis." Oncogene **25**(16): 2297-2303.

Xu, J., F. A. Ismat, T. Wang, J. Yang and J. A. Epstein (2007). "NF1 regulates a Ras-dependent vascular smooth muscle proliferative injury response." Circulation **116**(19): 2148-2156.

Zochodne, D. (1984). "Von Recklinghausen's vasculopathy." Am J Med Sci **287**(1): 64-65.

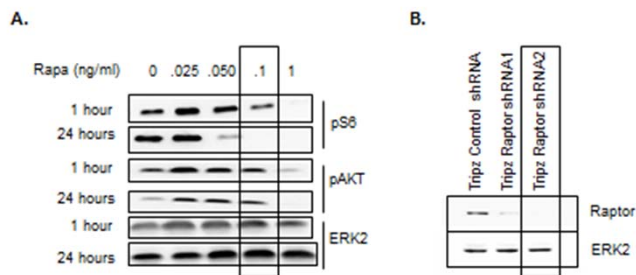


Figure 1. Rapamycin selectively inhibits mTORC1 low doses A) HUVECs were treated with various doses of rapamycin and cell lysates were probed for p-S6 and p-Akt at indicated times. B) Normalized expression of p-S6 and p-Akt reported as means of \pm standard error of three experiments.

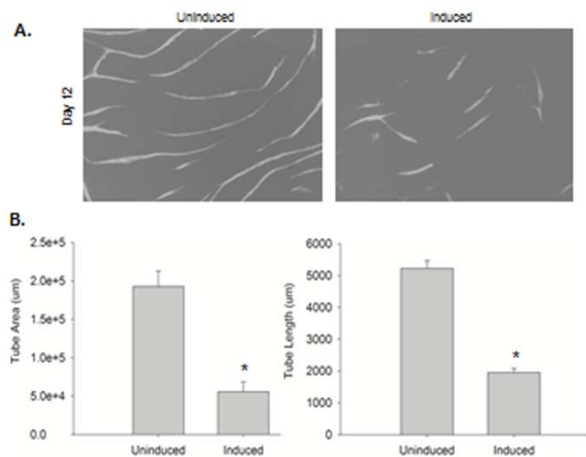


Figure 3. Raptor knockdown decreases growth of developing vessels A) *In vitro* co-culture assay showing endothelial cells expressing TripZ Raptor shRNA in either induced or uninduced conditions. Raptor knockdown was induced with doxycycline from day 0 and developed for 12 days. Images (gray scale, 8 bit) were taken at day 12 for both conditions. C) Total tube area and length in micrometers (μ m) from three similar experiments. Columns, reported as mean of \pm standard error of three experiments; bars, SE. *, $p < 0.01$.

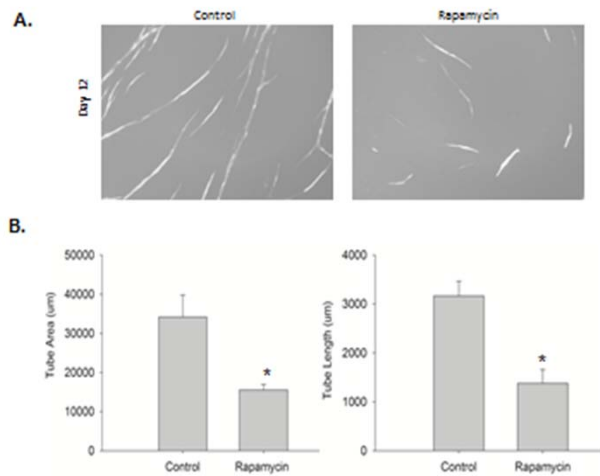


Figure 2 Selective inhibition of mTORC1 decreases growth of developing vessels. A) *In vitro* co-culture assay showing cells treated with rapamycin from day 0 and developed for 12 days. Endothelial cells expressing RFP were used for this assay. Images (gray scale, 8 bit) were taken at day 12 for both control and rapamycin treated conditions. B) Total tube area and length in micrometers (μ m) from three similar experiments. Columns, reported as means of \pm standard error of three experiments; bars, SE. *, $p < 0.01$.

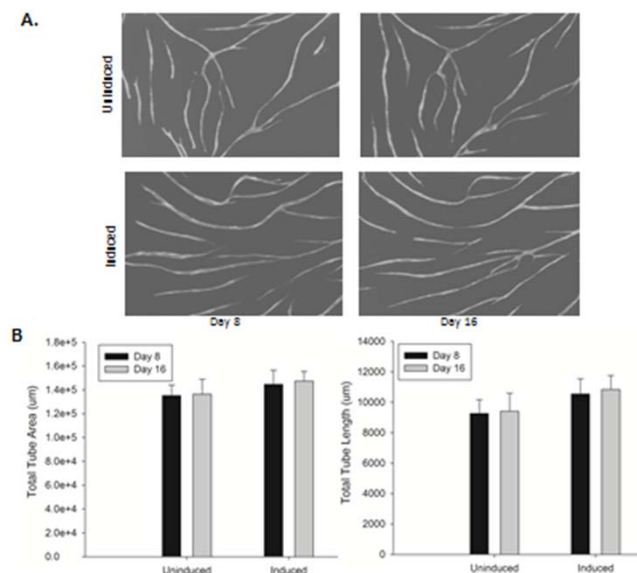


Figure 4 TORC1 inhibition does not affect established vessels. A) *In vitro* co-culture assay showing vessels developed in normal condition for 8 days and then treated with doxycycline to induce Raptor shRNA, along with untreated control. Endothelial cells expressing RFP were used for this assay to visualize vascular structures. Images (gray scale, 8 bit) from the same field were taken at day 8 and 16 for both control and Raptor knockdown. B) Total tube area and length in micrometers (μ m) from three similar experiments. Columns, reported as mean of \pm standard error of three experiments; bars, SE.

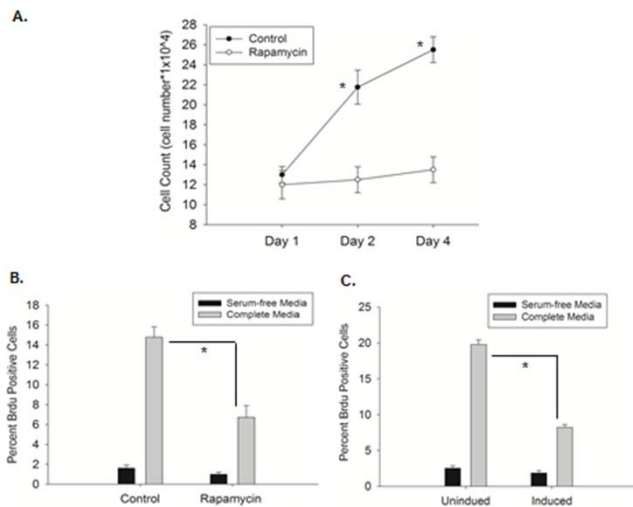


Figure 5 TORC1 is required for endothelial cell growth and proliferation. A) Endothelial cells at indicated times in complete media and counted after trypsinization at day 1, 2 and 4 using cell counter. Points, mean of triplicate determinations from three experiments; bars, SE. *, $p < 0.01$. B) Normalized percent BrdU positive cells in both MCDB serum free media and complete media for both conditions. (C) Identical experiment to that in (B) except TORC1 inhibition achieved by induction of Raptor shRNA. Columns, reported as mean of \pm standard error of three experiments; bars, SE. *, $p < 0.01$.

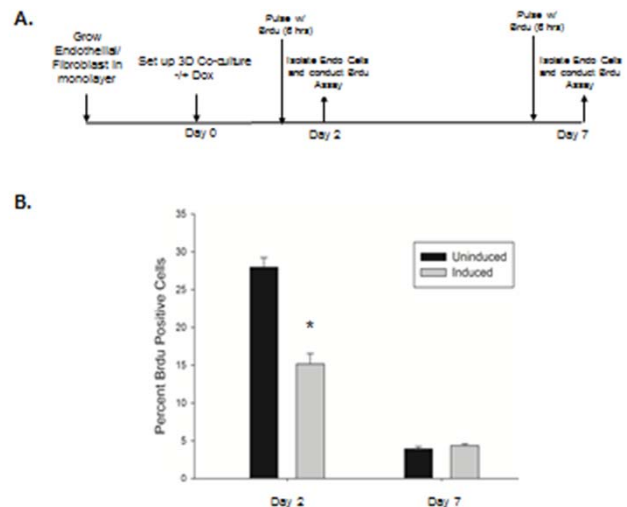


Figure 6 Raptor knockdown decreases endothelial cell proliferation in a co-culture model. A) A schematic showing timeline of the experiment presented in panel B and C. B) Normalized percent BrdU positive cells at day 2 and 7 for both conditions from three experiments. Columns, reported as mean of \pm standard error of three experiments; bars, SE. *, $p < 0.01$ at day 2.

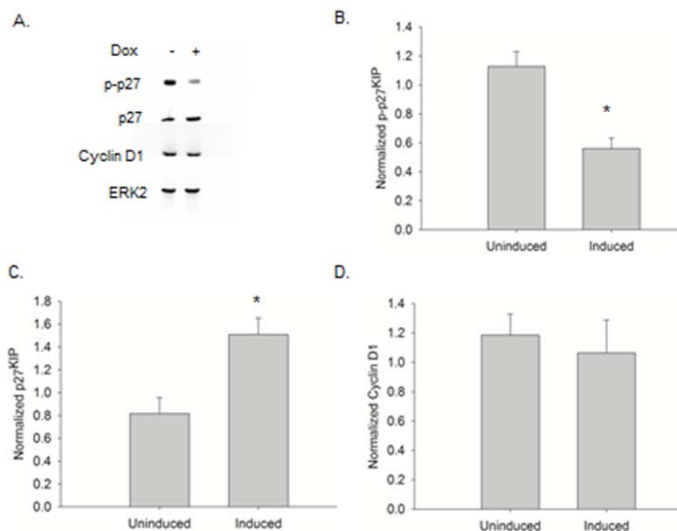


Figure 7 Selective mTORC1 inhibition decreases phosphorylation and accompanying stabilization of p27^{KIP} without affecting cyclin D1 level. A) Huvecs were treated with doxycycline to induce Raptor shRNA for 48 hours and cell lysates were probed for p-p27, p27 and cyclin D1. B-D) Normalized p-p27, p27 and cyclin D1 reported as means of \pm standard error of three experiments.

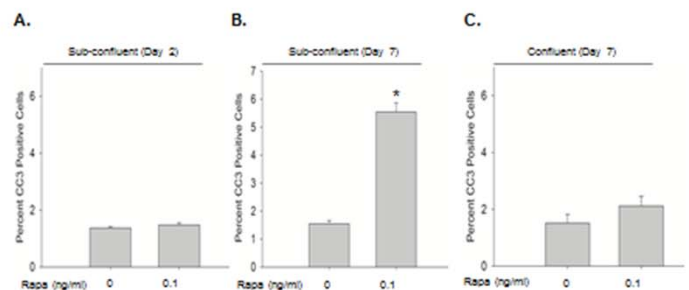


Figure 8 Rapamycin induces cleaved caspase 3 (CC3) in exponentially growing cells in a time dependent manner. A) Percent of cells positive for CC3 after rapamycin treatment for 2 days when cells were at a sub-confluent state. Columns, reported as mean of \pm standard error of three experiments; bars, SE. B) Percent of cells positive for CC3 after rapamycin treatment for 7 days when cells were at a sub-confluent state. Columns, reported as mean of \pm standard error of three experiments; bars, SE. *, $p < 0.01$ C) Percent of cells positive for CC3 when cells were treated with rapamycin for 7 days at a confluent state. Columns, reported as mean of \pm standard error of three experiments; bars, SE.

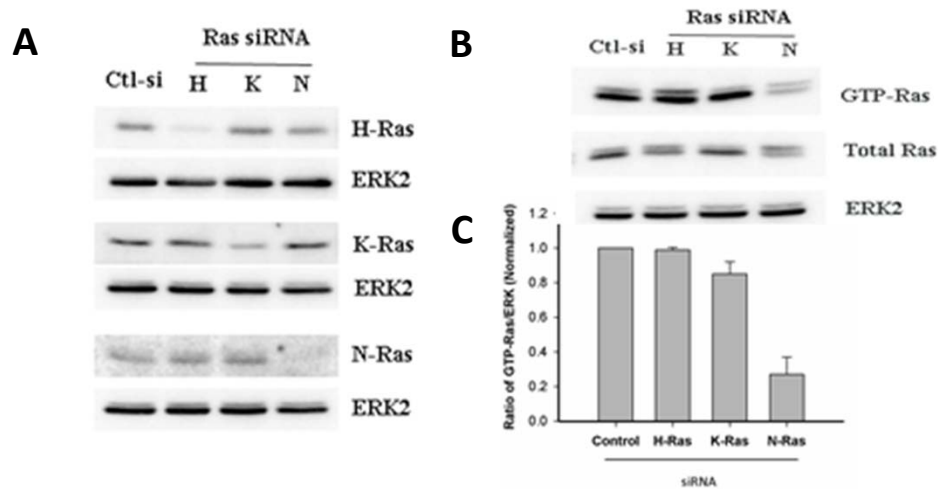


Figure 9. Knockdown of Ras isoforms reveals regulation of N-Ras by NF1 (A) HUVECs expressing NF1 shRNA were electroporated with isoform specific siRNA or control and evaluated for knockdown with isoforms specific antibodies of (B) the presence of active Ras using affinity pull-down assay. (C) Results from three experiments are normalized and quantified.

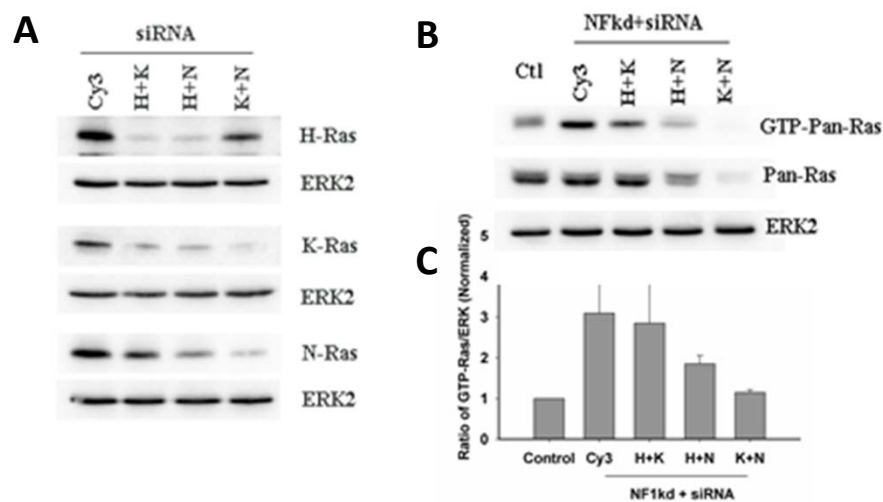


Figure 10. Double knockdown of Ras isoforms reveals regulation of N-Ras by NF1 (A) HUVECs expressing NF1 shRNA were electroporated with combinations of isoform specific siRNA or control and evaluated for knockdown with isoforms specific antibodies of (B) the presence of active Ras using affinity pull-down assay. (C) Results from three experiments are normalized and quantified.

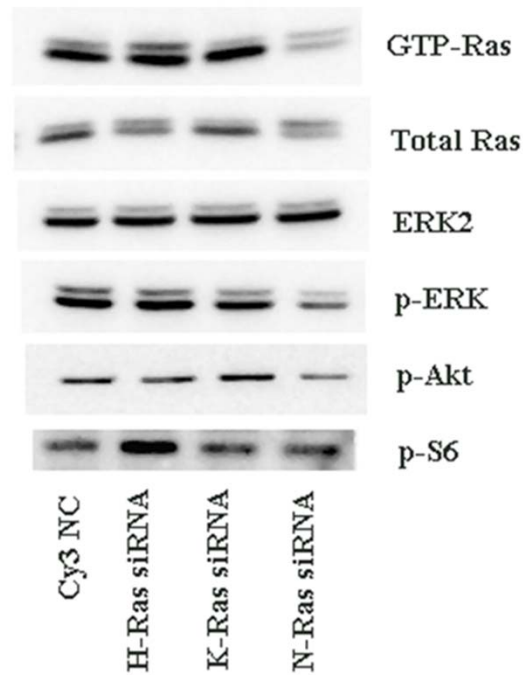


Figure 11. Knockdown of N-Ras ablates activation of signals induced by loss of NF1 (A) HUVECs expressing NF1 shRNA were electroporated isoform specific siRNA or control and evaluated for activation of Ras GTP and downstream effectors such as ERK, AKT and phospho-S6 using phospho-specific antibodies.

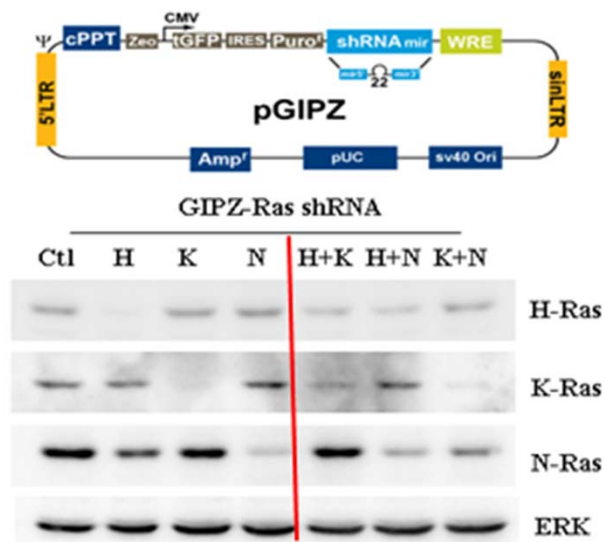


Figure 12 Stable knockdown of Ras isoforms in primary HUVECs (A) HUVECs were infected with lentiviruses coding for isoform specific shRNA or control and evaluated for knockdown with isoforms specific antibodies alone or in combination (right side of red line).

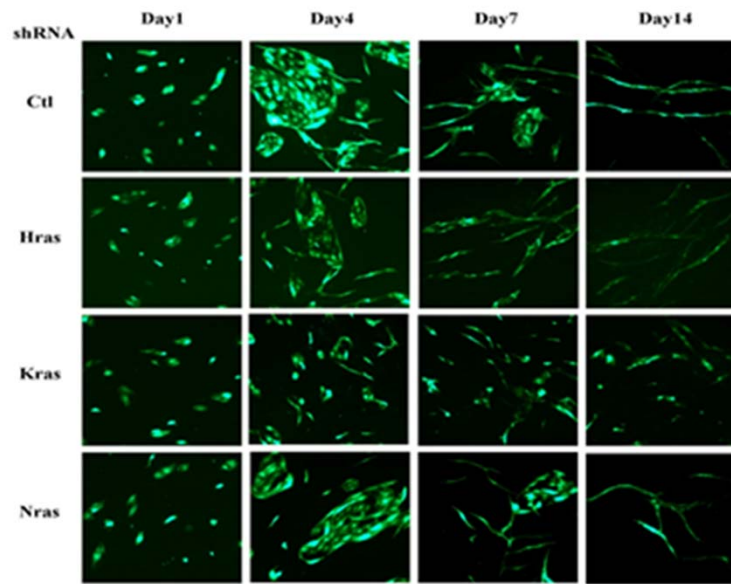


Figure 13. *K-Ras is required for vascular morphogenesis* Cells infected with lentiviruses designed with shRNA to knockdown Ras isoforms were analyzed using an endothelial-fibroblast co-culture assay for the times indicated.

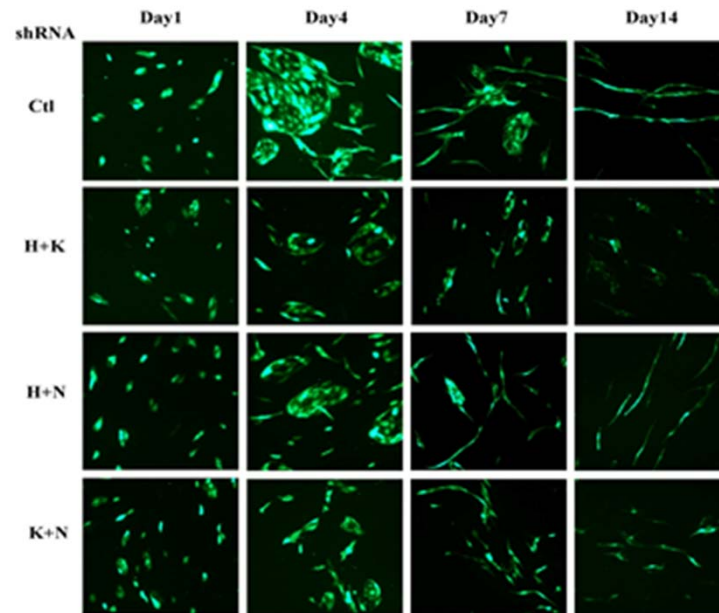


Figure 14. *K-Ras is sufficient for vascular morphogenesis* Cells were double infected with lentiviruses designed with shRNA to knockdown Ras isoforms were analyzed using an endothelial-fibroblast co-culture assay for the times indicated. Cells knocked with both H and N isoforms are still capable of forming tubes.

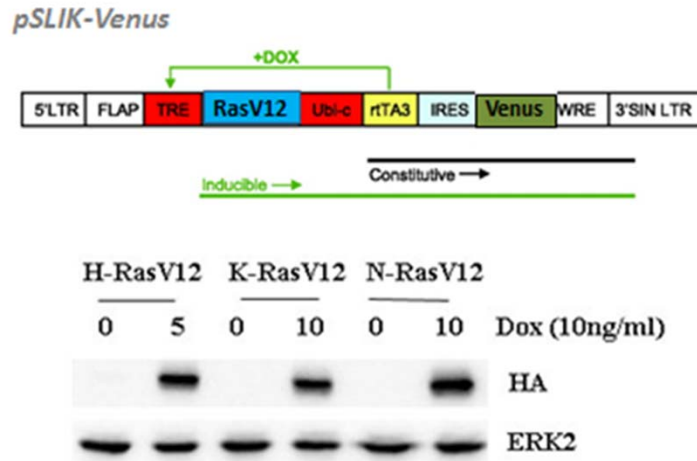


Figure 15. Inducible expression of activated isoforms of Ras. HUVECs were infected with lentiviruses designed to express specific Ras isoforms under doxycycline inducible control (Dox). All isoforms carry an activating mutation at valine 12 and all are epitope tagged to insure equal expression across isoforms.

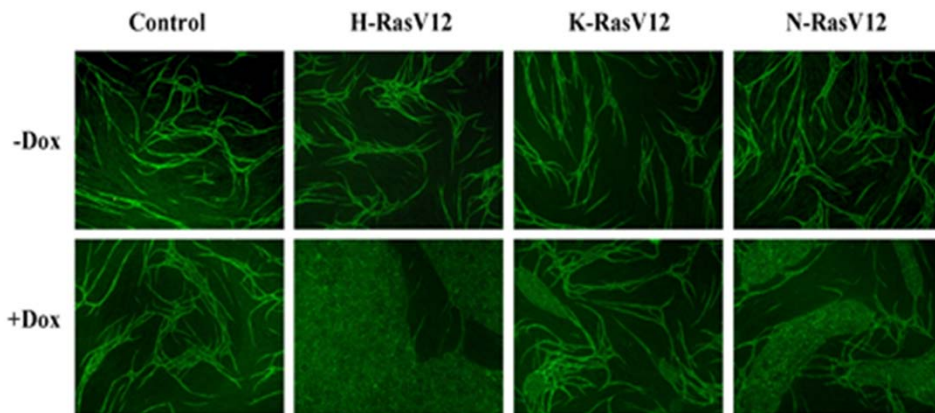


Figure 16. H-Ras preferentially alters vascular morphogenesis. HUVECs were infected with lentiviruses designed to express specific Ras isoforms under doxycycline inducible control (Dox). Cells were placed in co-culture with primary fibroblasts and induced with doxycycline to express the mutant Ras or left uninduced for 14 days. Detection of vascular structures is by labeling with FITC-UEA-1 lectin. Micrographs were taken at 40X magnification.

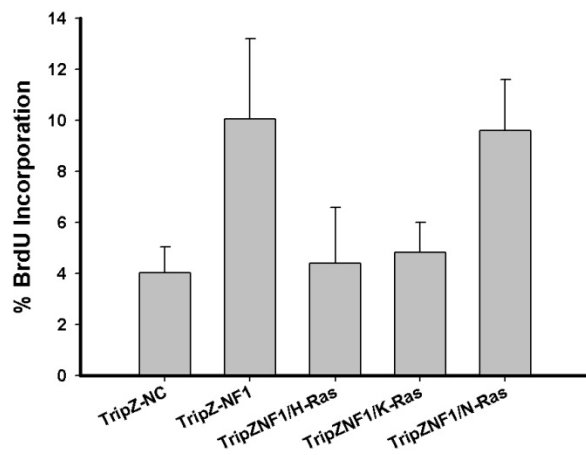


Figure 17. Effect of Isoform knockdown on NF1-mediated proliferation. HUVECs were double-infected with lentiviruses designed to express shRNA targeting specific Ras isoforms under doxycycline inducible control (Dox) in combination with knockdown of NF1. Cells were serum starved and analyzed for BrdU incorporation.

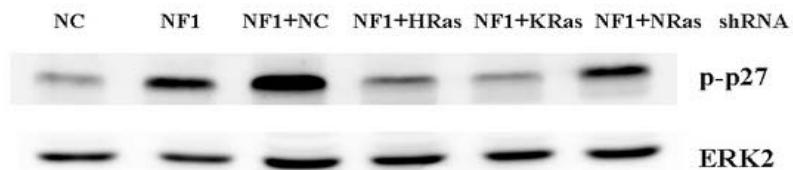


Figure 18. Effect of Isoform knockdown on NF1-mediated p27-phosphorylation.

HUVECs were double-infected with lentiviruses designed to express shRNA targeting specific Ras isoforms under doxycycline inducible control (Dox) in combination with knockdown of NF1. Cells were western blotted for phosphoP-27 and ERK2 as a loading control.

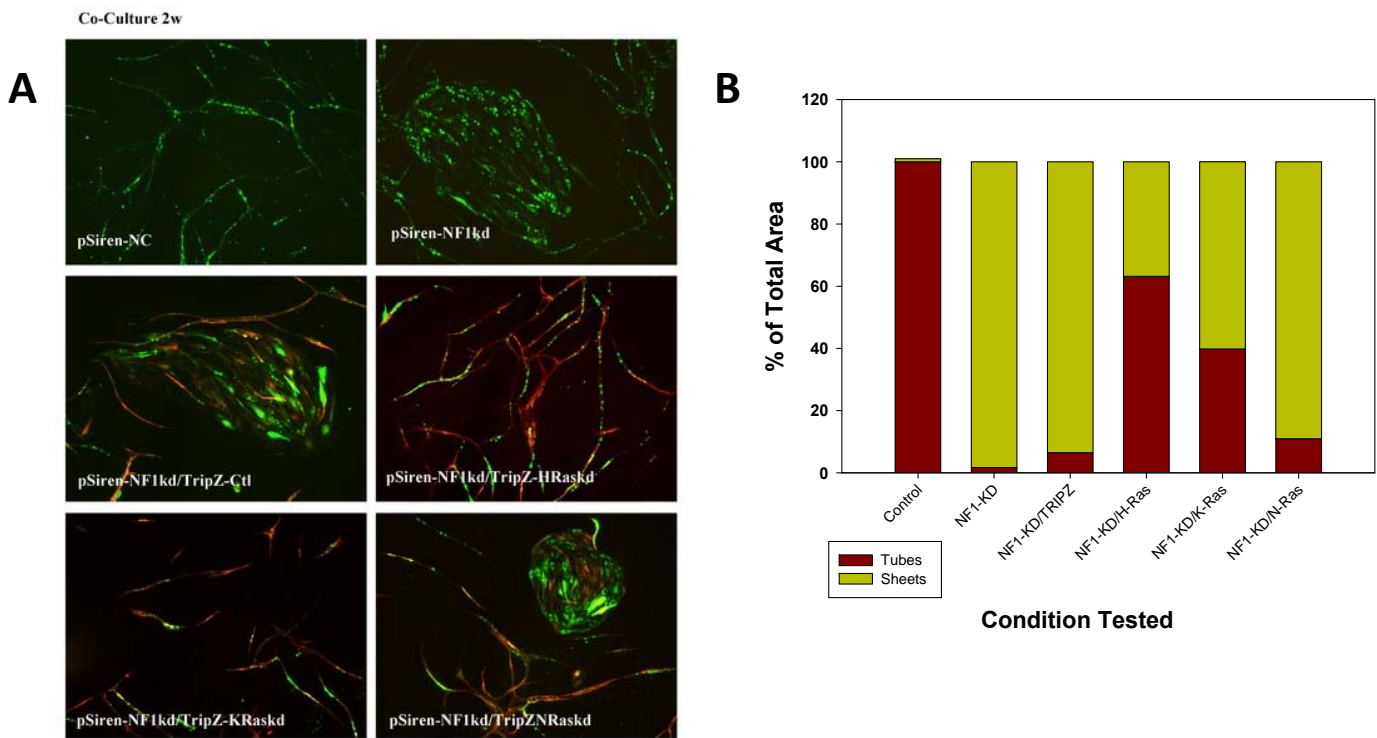


Figure 19. Effect of Isoform knockdown on NF1-mediated proliferation. HUVECs were double-infected with lentiviruses designed to express shRNA targeting specific Ras isoforms under doxycycline inducible control (Dox) in combination with knockdown of NF1. Cells were placed in co-culture with primary fibroblasts and induced with doxycycline to express the mutant Ras for 14 days. (A) Detection of vascular structures NF1 shRNA is linked to GFP expression, Ras isoform to RFP expression. Images were acquired on both channels and merged. (B) Images were blindly quantified for the presence of tube like structures and sheets and represented by a stacked bar graph.

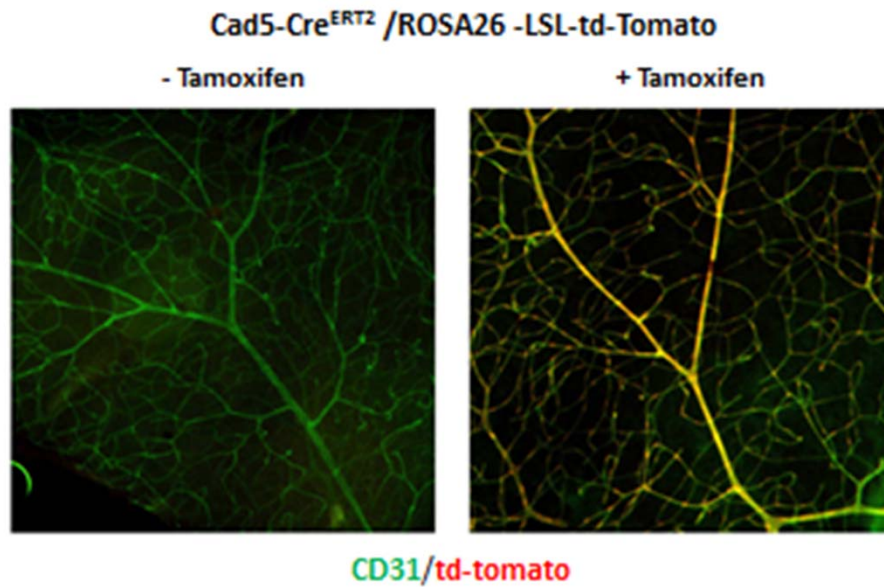


Figure 20 Endothelial Specific, Tamoxifen-inducible CRE expression. Cad5-Cre^{ERT2} /ROSA26 -LSL-td-Tomato mice were used to test penetrance and specificity of endothelial specific cre expression. Induction with Tamoxifen for 3 days was followed by a waiting for 2 days and then sacrifice. Retinal whole mounts were analyzed by CD31 staining to detect all endothelial cells and red fluorescence from the Cre-responsive reporter, td-Tomato.

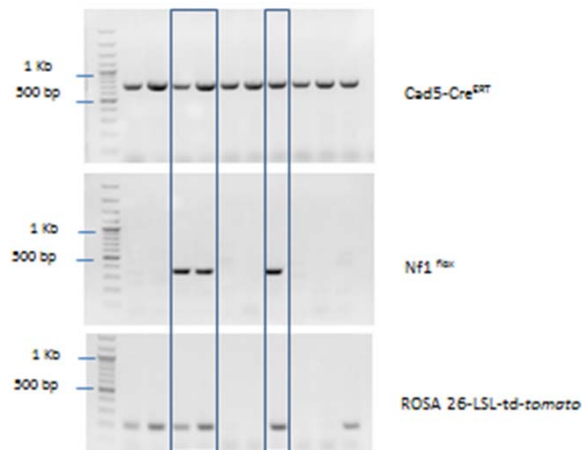
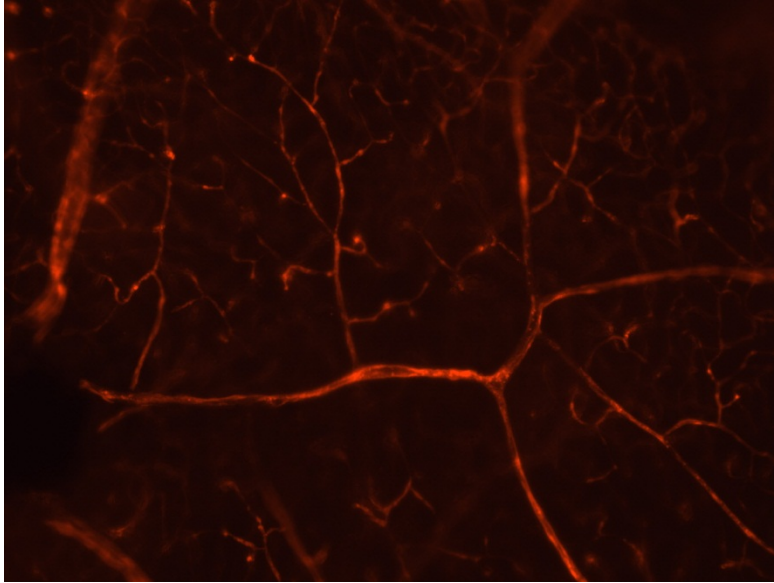


Figure 21 Generation of Endothelial Cell Specific, NF1 Deletion Strain. Endothelial specific Cad5-CreERT was mated with NF1 flox and ROSA26 -td-Tomato mice to generate mice carrying all three alleles, as highlighted by the blue boxes. These mice will be used to breed with similarly produced mice with an NF1 +/- background for the proposed experiments.

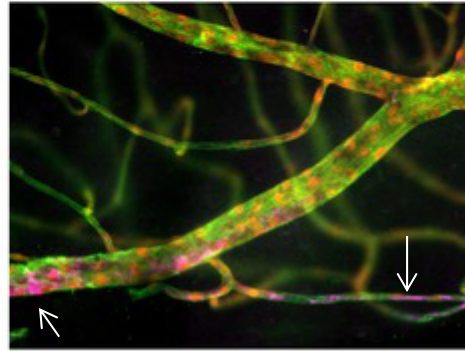
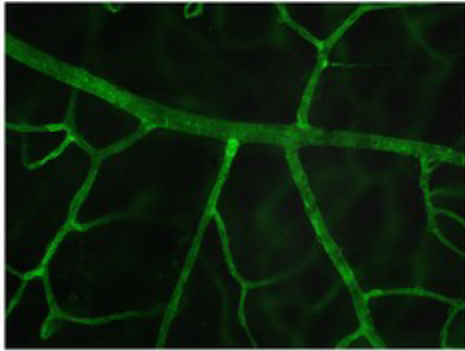
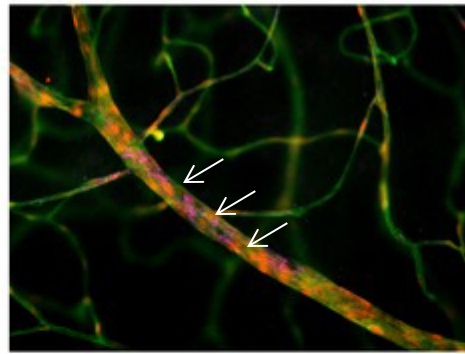
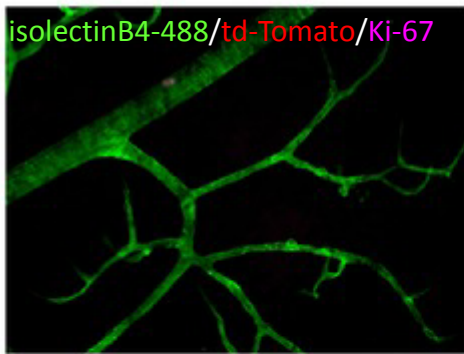
Cad5-Cre^{ERT2}/Rosa26-LSL-td-Tomato/NF1^{flox/flox}

A



Tamoxifen

B



Uninduced

Tamoxifen

Figure 22 Potential vascular abnormalities upon loss of *NF1* in the vascular endothelium. Endothelial specific *Cad5-CreERT* was mated with *NF1-flox* and *ROSA26-td-Tomato* mice to generate mice carrying all three alleles. These mice were induced with tamoxifen, as indicated, to activate Cre activity. As shown in Panel A, Cre activity is highly penetrant, activating Tomato expression in nearly all of the vascular endothelial cells. Panel B shows mice treated with vehicle or tamoxifen and sacrificed 30 days later. Retinal whole mounts were analyzed with a vascular endothelial marker (isolectin B4), and a marker for proliferation (Ki67), as well as for expression of tomato. Arrows indicate positive Ki-67 staining in endothelial cells.

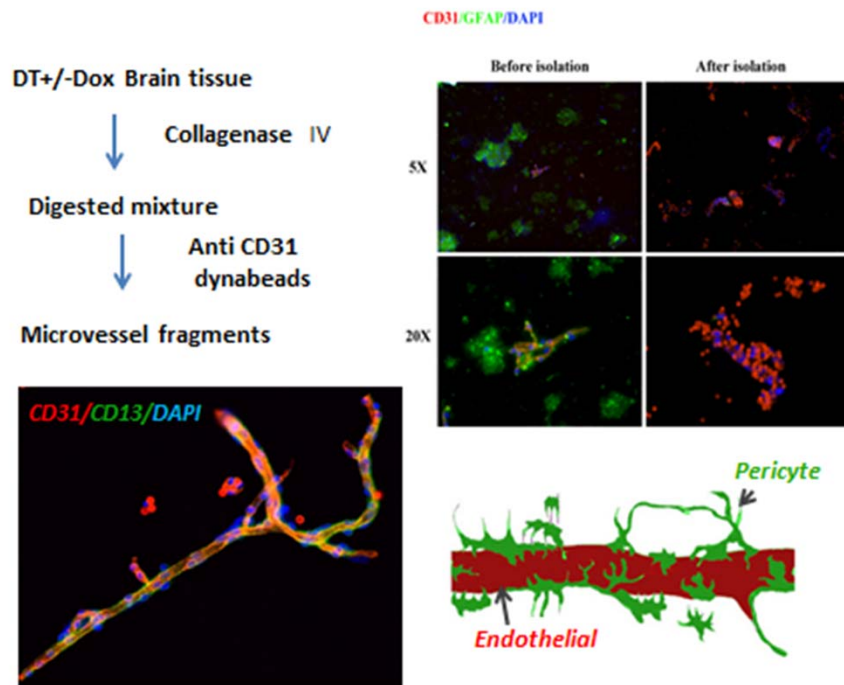


Figure 23 Isolation of microvessel fragments from brain Using a combination of selective digestion and immunoisolation we are to separate intact microvessel fragments away from neurons, astrocytes (GFAP) and microglial cells of the brain. This permits biochemical analysis of signaling and gene transcription.

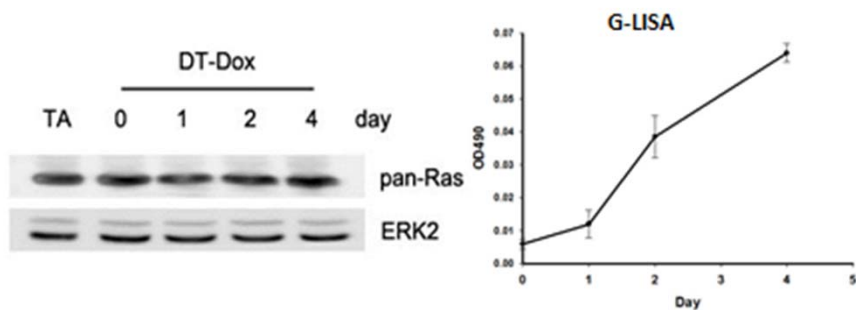


Figure 24 Activation of Ras in microvessel fragments from brain We prepared microvessel fragments from control mice or from bi-transgenic mice expressing activated Ras only in the endothelium upon dox removal. Doxycycline was removed from the diet for the days indicated and microvessel fragments were isolated as indicated above. Microvessels were lysed and analyzed for Ras activation using a G-lisa assay, an affinity assay for GTP-Ras in an ELISA format.

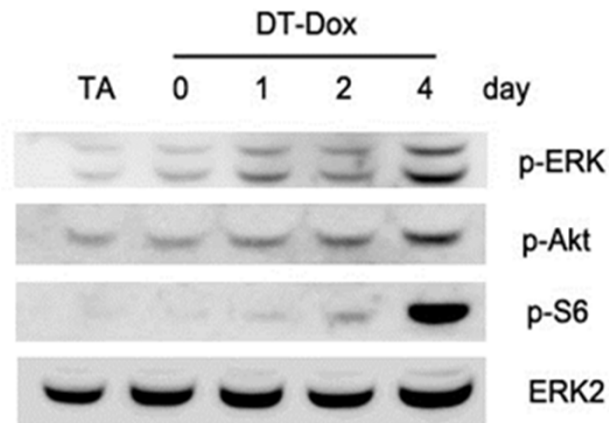


Figure 25 Activation of Ras signal transduction in microvessel fragments from brain We prepared microvessel fragments from control mice or from bi-transgenic mice expressing activated Ras only in the endothelium upon dox removal. Doxycycline was removed from the diet for the days indicated and microvessel fragments were isolated as indicated above. Microvessels were lysed and analyzed for the presence of active Ras-related signaling using phospho-specific antibodies. ERK2 serves as a lysate loading control.

Loss of NF1 Expression in Human Endothelial Cells Promotes Autonomous Proliferation and Altered Vascular Morphogenesis

Anshika Bajaj¹, Qing-fen Li¹, Qingxia Zheng, Kevin Pumiglia*

Center for Cell Biology and Cancer Research, Albany Medical College, Albany, New York, United States of America

Abstract

Neurofibromatosis is a well known familial tumor syndrome, however these patients also suffer from a number of vascular anomalies. The loss of NF1 from the endothelium is embryonically lethal in mouse developmental models, however little is known regarding the molecular regulation by NF1 in endothelium. We investigated the consequences of losing NF1 expression on the function of endothelial cells using shRNA. The loss of NF1 was sufficient to elevate levels of active Ras under non-stimulated conditions. These elevations in Ras activity were associated with activation of downstream signaling including activation of ERK, AKT and mTOR. Cells knocked down in NF1 expression exhibited no cellular senescence. Rather, they demonstrated augmented proliferation and autonomous entry into the cell cycle. These proliferative changes were accompanied by enhanced expression of cyclin D, phosphorylation of p27^{KIP}, and decreases in total p27^{KIP} levels, even under growth factor free conditions. In addition, NF1-deficient cells failed to undergo normal branching morphogenesis in a co-culture assay, instead forming planar islands with few tubules and branches. We find the changes induced by the loss of NF1 could be mitigated by co-expression of the GAP-related domain of NF1 implicating Ras regulation in these effects. Using doxycycline-inducible shRNA, targeting NF1, we find that the morphogenic changes are reversible. Similarly, in fully differentiated and stable vascular-like structures, the silencing of NF1 results in the appearance of abnormal vascular structures. Finally, the proliferative changes and the abnormal vascular morphogenesis are normalized by low-dose rapamycin treatment. These data provide a detailed analysis of the molecular and functional consequences of NF1 loss in human endothelial cells. These insights may provide new approaches to therapeutically addressing vascular abnormalities in these patients while underscoring a critical role for normal Ras regulation in maintaining the health and function of the vasculature.

Citation: Bajaj A, Li Q-f, Zheng Q, Pumiglia K (2012) Loss of NF1 Expression in Human Endothelial Cells Promotes Autonomous Proliferation and Altered Vascular Morphogenesis. PLoS ONE 7(11): e49222. doi:10.1371/journal.pone.0049222

Editor: Salvatore V. Pizzo, Duke University Medical Center, United States of America

Received: May 24, 2012; **Accepted:** October 4, 2012; **Published:** November 7, 2012

Copyright: © 2012 Bajaj et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: Funding was provided by award W81XWH-09-1-0432 from the Neurofibromatosis Research Program of CDMRP (<http://cdmrp.army.mil/nfrp/default.shtml>) and by the generosity of the David E. Bryant Trust. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: pumiglk@mail.amc.edu

¶ These authors contributed equally to this work and are listed in alphabetical order.

‡ Current address: GE Global Research, Niskayuna, New York, United States of America

Introduction

Mutations in the NF1 gene cause Neurofibromatosis type 1, an autosomal dominant disease that affects approximately 1 in 3000 individuals, making it one of the most common inherited genetic disorders [1]. NF1 has variable clinical manifestations. Most commonly observed changes include alterations in skin pigmentation (café au-lait spots as well as freckling) and the presence of benign and malignant nerve sheath tumors termed neurofibromas [1]. Importantly, a significant clinical manifestation of NF1 disease includes vascular disease. Patients with NF1 disease make up a significant portion of all those patients presenting with renal artery stenosis and early-onset cerebral vascular disease [2] and cardiovascular disease is a significant contributor to premature death in NF1 patients, particularly among younger patients. One study suggested that vasculopathy was over seven times more likely to occur in NF1 patients under 30 compared to their unaffected peers [3].

NF1 is clinically associated with a pleiotropic array of vascular abnormalities including stenosis, malformations, aneurysms, and hypertension. As a consequence these patients show a markedly elevated risk of cerebrovascular accidents [2]. Previous studies in mice have suggested an important role of smooth muscle cells [4] and bone marrow cells [5] in neointimal hyperplasia, inflammation and exaggerated response to injury including enhanced angiogenesis. However little is currently known about factors contributing to vascular malformations and the role of endothelial cells in regulating these changes. In addition, the endothelium is critically poised to regulate blood vessel formation, vascular tone, inflammation, as well as coagulation, thus a better understanding of the role of NF1 in regulating the function of the vascular endothelium may be critical to understanding many facets of this disease and reducing its morbidity.

Previous studies support the notion that NF1 has a critical role in the vascular endothelium. Deletion of NF1 from the vascular

endothelium results in embryonic lethality [6] and NF1 haploinsufficient mice show exaggerated angiogenic responses [7]. As NF1 is a Ras-GTPase activating protein, changes in Ras activation are often associated with the loss of NF1 and data have been published suggesting that shRNA mediated knockdown of NF1 can augment growth factor mediated Ras activation and downstream signaling in endothelial cells [8]. NF1 is a large protein however, with other signaling effects, including changes in cAMP and mTOR, which can be Ras-independent [9]. We have recently published that activation of Ras in primary endothelial cells is sufficient to drive a pro-survival, pro-proliferative phenotype that disrupts normal vascular morphogenesis [10]. It is unclear if loss of NF1 is sufficient to enhance basal activation of Ras and initiate cellular responses in the absence of additional growth factor signaling. We conducted these studies to determine if the loss of NF1 is sufficient to initiate cellular signaling and alter endothelial cell function, to determine the role of Ras and other cellular signals acting downstream of NF1, and to evaluate how these changes affect the behavior of endothelial cells in a complex microenvironment.

Results

Knockdown of *NF1* in Primary Endothelial Cells Activates Ras and Downstream Signaling

To generate endothelial cells lacking neurofibromin a shRNA against NF1 was cloned into pSIREN-RetroQ-ZsGreen retroviral expression vector, as previously described [8] allowing stable knockdown of neurofibromin both at the protein (Fig. 1A) and mRNA levels (not shown). Primary cells have previously demonstrated growth arrest in response to the loss of NF1 [11]. To minimize potential confounding senescence effects, to regulate expression, and also to safeguard against RNAi mediated off-target effects, a different NF1 knockdown sequence was cloned into a lentiviral pTRIPZ vector that permits inducible knockdown of the protein in the presence of 0.5 μ g/mL doxycycline whereas no knockdown was seen in the absence of doxycycline (Fig. 1B). Using both types of cells we examined the levels of active Ras in quiescent cells. Knockdown of NF1 in human endothelial cells using the inducible miR-based shRNA results in increased levels of active Ras under basal conditions (Fig. 1C). Similarly, stable knockdown of NF1 also results in enhanced levels of Ras-GTP (Figure S2), even following several passages suggesting limited down-regulation or cellular compensation of this response. The enhanced levels of Ras-GTP were sufficient to trigger downstream signaling, as knockdown of NF1 resulted in enhanced activation of ERK, PI-3'-Kinase/AKT (Fig. 1D), even in the absence of any added growth factors. Another important signaling network linked to tumors and cells lacking NF1 is the mTOR signaling pathway [12,13]. It has been shown that activation of mTOR pathway in nerve sheath tumor cell lines is essential for neurofibroma formation [14]. As the role of NF1 in regulating mTOR signaling in endothelial cells is unknown, we investigated this by examining the phosphorylation of ribosomal S6, a substrate of the TORC1 activated kinase, S6K. We found loss of NF1, even the absence of added growth factors, sufficient to stimulate mTORC1/S6 signaling. Similar effects were observed with both the inducible and the stable knockdown, suggesting little temporal or kinetic differences between the constructs or approaches (Figs. 1D and S2). These data provide evidence that knockdown of the Ras-GAP NF1 in human endothelial cells is sufficient to elevate cellular GTP-Ras levels and activate downstream signaling in the absence of added growth factors.

Cellular Proliferation in Endothelial Cells Lacking NF1

Down-regulation of signaling is part of a feedback response that results in replicative arrest in primary fibroblasts. However, we see no evidence of signal dampening in endothelial cells lacking NF1 even after prolonged culturing. To determine if the chronic activation of Ras and the related signaling results in replicative senescence, we examined several aspects of cellular growth. Initially, we evaluated population doublings, often used to monitor oncogene-induced senescence. In contrast to the effects observed following NF1-loss in fibroblasts [11], we found no signs of growth arrest in NF1-knockdown cells, rather they showed enhanced proliferative capacity (Fig. 2A). Given the enhanced long-term proliferative capacity of NF1-knockdown cells and the previous report that NF1-knockdown could enhance VEGF-induced proliferation, we next sought to determine if NF1 was sufficient to induce endothelial cell proliferation in the absence of growth factors. As shown in Fig. 2B, even in the absence of added mitogenic factors, endothelial cells lacking NF1 expression demonstrated enhanced entry into S-phase of the cell cycle as measured by BrdU incorporation. The enhanced proliferation was accompanied by upregulation of cyclin D levels, enhanced phosphorylation of p27KIP, and a corresponding decrease in levels of total p27 (Fig. 2C). Thus, the signaling induced by the loss of NF1 is sufficient to stimulate DNA synthesis as a consequence of cyclin D induction and the loss of the Cyclin-dependent kinase inhibitor, p27KIP.

Loss of NF1 is Associated with Altered Vascular Morphogenesis

NF1 patients are known to have several distinct types of vascular anomalies associated with the disease, including vascular malformations. At this point it is unclear if this is the result of effects arising in endothelial cells or as a consequence of an altered and pro-angiogenic microenvironment. The altered proliferative control seen upon the loss of NF1 might contribute to altered morphogenic responses by human endothelial cells. To test this we utilized a co-culture assay where human endothelial cells in the presence of human primary fibroblasts will typically proliferate for a cycle or two followed by a cessation of proliferation and the formation of branched networks of endothelial cell tubules that contain patent lumens [10,15,16]. When we performed this assay we found that endothelial cells lacking NF1 had an altered morphogenic response. In lieu of forming elongated and branched networks, these cells tended to form planar, sheet like structures which showed few elongations and branches, as if an essential signal to “differentiate” into vascular structures was compromised (Fig. 3). Identical phenotypes were seen whether we used constitutive shRNA (pSiren) knockdown or the inducible miR-based construct (TripZ).

Effects on Endothelial Cells Following the Loss of NF1 are a Consequence of Ras Activation

The loss of NF1 is sufficient to activate cellular Ras. However, NF1 is a large protein with several alternative signaling paradigms through which it can affect cell function in a Ras-independent manner, including changes in cAMP signaling [9,17]. Therefore, we sought to determine if the effects we were seeing were the result of Ras activation. To test this we co-infected endothelial cells with inducible lentiviral vectors to knockdown NF1 and then re-express the GAP-related domain of NF1 (GRD) in order to “rescue” the Ras GAP functions of NF1 (this domain is outside the targeted region). Purified cells were obtained by two-color sorting (Red; TRIP-Z (NF1-KD) co-expressed RFP; Green; pSLIK (GRD) co-

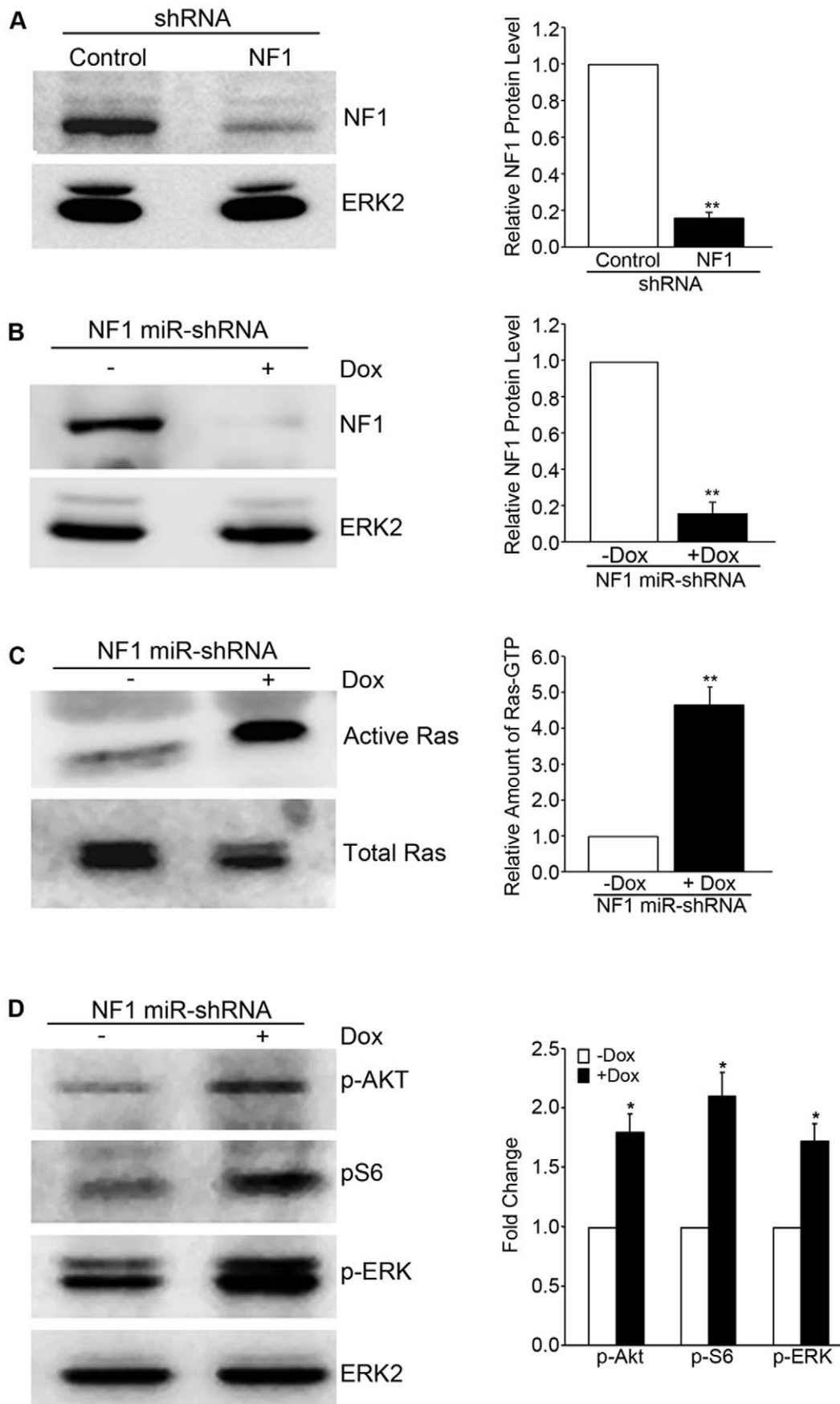


Figure 1. Knockdown of NF1 in Human Endothelial Cells is Sufficient to Activate Ras and Induce Cellular Signaling. (A) Early passage HUVECs were infected with a pSIREN-GFP retroviral vector carrying either a control or a NF1 shRNA and sorted for GFP expression. Western blot analysis confirmed knockdown of neurofibromin. (B) Early passage endothelial cells were infected with a pTRIPZ lentiviral vector expressing either a non-silencing control or a NF1 miR-based shRNA. The infected cells were induced with 1 μ g/mL doxycycline for 48 h to induce expression of the microRNA along with red fluorescent protein (expressed in tandem) and sorted for RFP expression. Western blot confirms knockdown of neurofibromin in the presence of doxycycline while no knockdown was seen in the uninduced cells. (C) Uninduced (–Dox) and induced (+Dox) endothelial cells were serum starved for 24 h and levels of active Ras (RasGTP) were determined by using GST-Raf pull-down assay (Pierce), according to the manufacturer's protocol. Total Ras in the total cell lysates confirmed similar amounts of protein input and was used to normalize GTP-Ras quantification. (D) Cells were treated as described above but western analysis was performed to measure activation of several key signaling proteins including phospho-Akt, phospho-S6, and phospho-ERK. Equal lysate loading was confirmed by monitoring total ERK2 levels. All experiments were performed in at least three independent sets of control and NF1 knockdown HUVECs and quantification results were averaged. (Identical results were seen with both knockdown vectors (see for example figure 6) Error bars represent standard error of the mean (** $p < 0.01$, * $p < 0.05$). doi:10.1371/journal.pone.0049222.g001

expressed YFP). As both vectors are tet-inducible, induction of knockdown also induces the “rescue” in the cells co-infected with the GRD expressing virus. As shown in Fig. 4, the expression of the GRD reverses the Ras activation seen following the loss of NF1, returning it to basal levels. This is accompanied by a similar reversal of the autonomous proliferation (Fig. 4B) that the loss of NF1 promotes. Expression of GRD also reduced the enhanced phosphorylation of p27 that is seen following the loss of NF1 with a coincident stabilization of this protein (Fig. 4C).

We also used these cells to determine if Ras activity was responsible for the hyperplastic morphogenic responses we observed. As shown in Fig. 5A, co-expression of the GRD reverses

the abnormal morphogenesis, with cells forming nicely branched and elongating networks when the GRD domain of Ras is co-expressed. These same cells were used to explore the reversibility of this phenotype. As shown in Fig. 5A, if the NF1-kD cells are allowed to form the hyperplastic structures for 14 days (a time at which normal endothelial cell networks have stabilized) and then the Dox is removed (to cease expression of the silencing shRNA), normal looking tubular branching structures emerge over the next 14 days. We also performed the converse experiment, where normal vessel-like structures were allowed to form and stabilize over 14 days (Fig. 5B). As Dox was added to induce the knockdown of NF1, vessel-like structures begin to thicken and fuse,

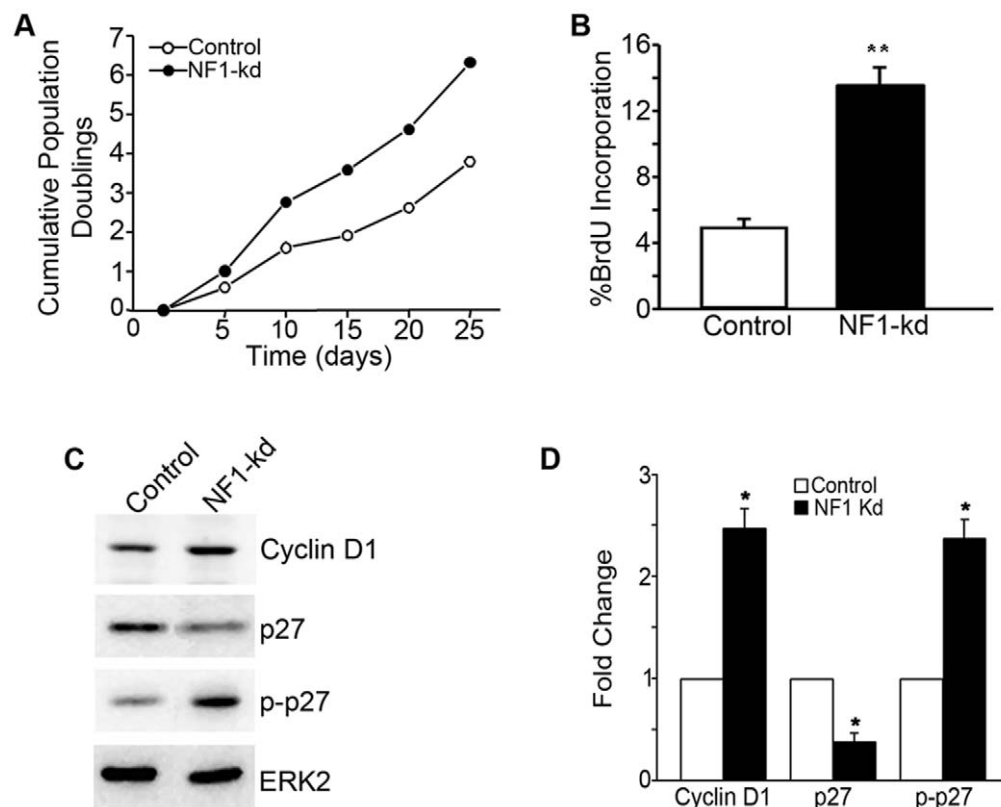


Figure 2. Loss of NF1 expression is sufficient to induce endothelial cell proliferation. (A) pSIREN Control and NF1 knockdown primary endothelial cells were grown in complete media under sub-confluent conditions and cumulative population doublings were recorded over the course of 25 days as described [10]. (B) Control or NF1 knockdown endothelial cells were serum deprived for 24 h prior to cells being pulsed with BrdU for 3 h. BrdU positive cells were quantified and the data was graphed as the % positive cells compared to total cell number. Data are presented as the mean with error bars representing the standard error (** $p < 0.01$). (C) In parallel to the experiments performed in (B), serum starved cell lysates were made after 24 h and probed with antibodies against cyclin D1, p27, phospho p27, and ERK2. ERK2 is used to insure equal loading of lysate. (D) In three independent experiments similar to those performed in (C), results were quantified and data expressed as fold-change from control. Error bars represent standard error of the mean (* $p < 0.05$). doi:10.1371/journal.pone.0049222.g002

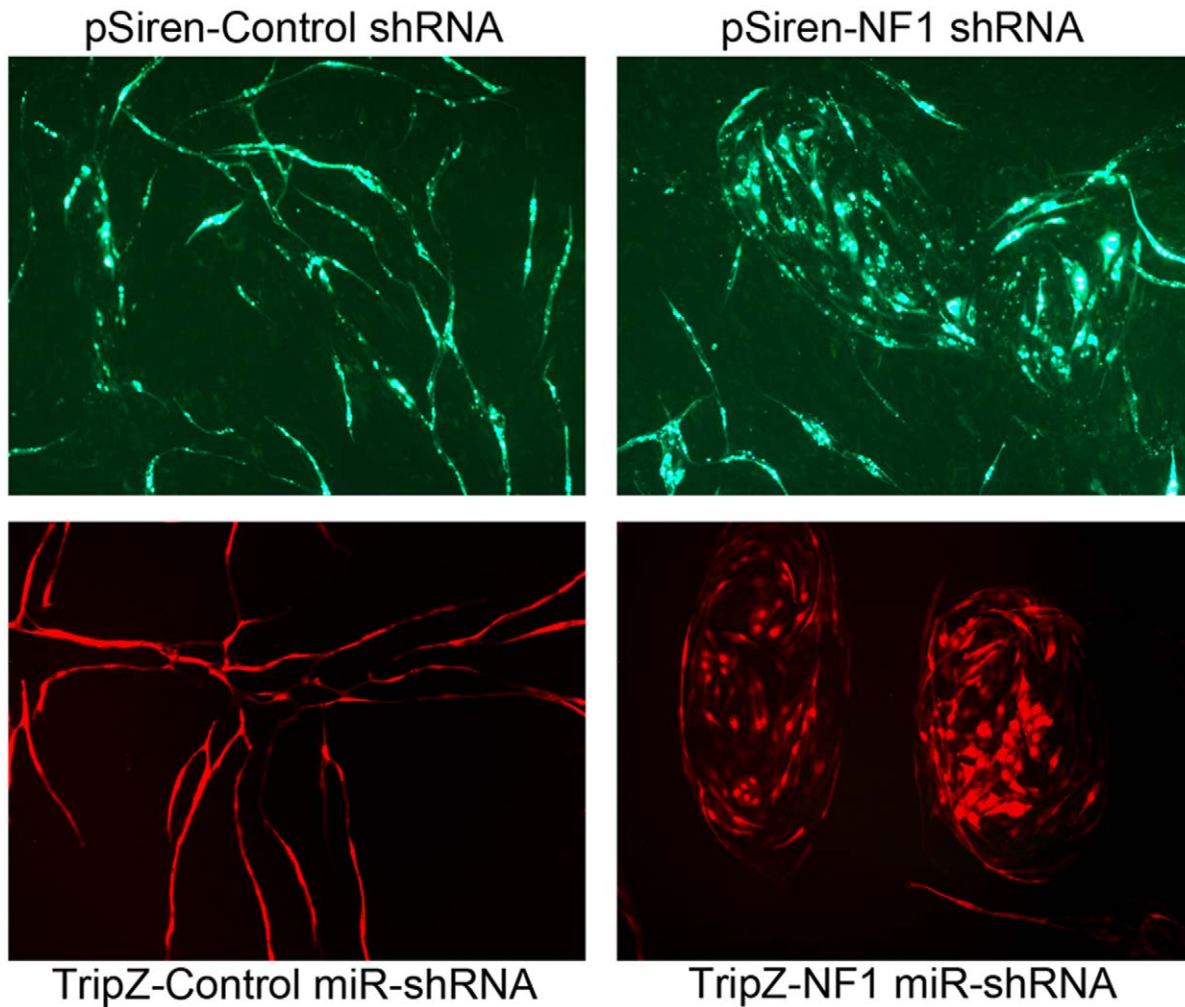


Figure 3. Loss of NF1 expression results in abnormal vascular morphogenesis. *Top Panel* - Primary endothelial cells infected with non-targeting pSIREN lentivirus (Control) or one directed toward knockdown of NF1 (NF1-KD). These cells were plated with primary fibroblasts in an admixed co-culture. After 14 days, endothelial cells were visualized by the vector expressed GFP. *Bottom Panel* - Endothelial cells infected with TRIPZ-Control or NF1miR-shRNA, were plated in co-culture with primary fibroblasts in the presence of doxycycline to induce expression of shRNA and RFP. Vascular structures were visualized at day 14 using RFP co-expressed in the endothelial cells.
doi:10.1371/journal.pone.0049222.g003

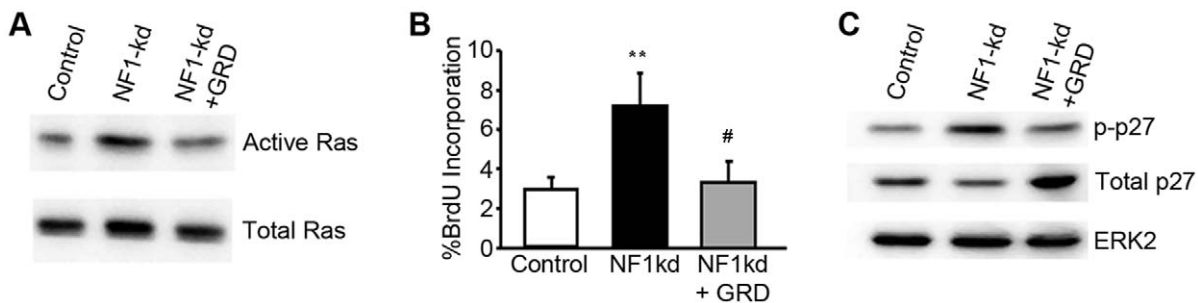


Figure 4. Enhanced proliferation following the loss of NF1 is a consequence of Ras activation. HUVECs were double infected with vectors coding for inducible NF1 knockdown shRNA and inducible expression of the GAP-related domain (GRD) of NF1 or empty vector. Cells were incubated for 24 h in the presence of 0.5 μ g/ml doxycycline to induce expression of NF1 shRNA as well as the co-infected cDNA (GRD or empty). Control cells are cultured in the absence of doxycycline to suppress expression of and cDNA. After 24 h in the presence or absence of doxycycline, cells were switched to serum-free medium. (A) Cells were analyzed for the presence of active Ras using GST-pull down. Total Ras was used to insure equivalent protein in the input lysates. (B) BrdU incorporation was measured 24 h after changing to serum and growth factor free conditions. Data represents the averages from triplicate determinations and error bars represent standard error of the mean. (** $p < 0.01$ compared to Control; # $p < 0.01$ compared to NF1-kd, (C) Lysates prepared under the conditions described above were probed for changes in phosphorylation of p27 and total p27 levels by western blotting. ERK was monitored as a loading control. All experiments were replicated in an independently generated set of doubly infected cells.
doi:10.1371/journal.pone.0049222.g004

suggesting that malformed vascular structures can arise, even from quiescent endothelial cells. In cells “rescued” by coordinate expression of the GRD, vessel morphology remained intact and unchanged throughout the experimental manipulations. Collectively these data strongly argue that maintenance of appropriate levels of Ras activation are critical for vascular morphogenesis both in developing vessels and in established vascular networks.

mTORC1 Activity is Essential for Endothelial Cell Proliferation and Abnormal Morphogenesis Following the Loss of NF1

We next sought to determine the mechanisms required for the autonomous proliferation seen in response to the loss of NF1 and the accompanying activation of Ras. Given the emerging role of mTOR-related signaling in other aspects of NF1 disease, we were interested in the role of this enzyme [18,19,20]. We used the minimal concentrations of inhibitors required to effectively inhibit Ras-related signaling back to basal levels and evaluated the effects on cellular signaling. We found we could effectively inhibit ERK activation with no significant effects on the S6 activation or AKT activation (Fig. 6A). Proliferation was completely inhibited by inhibition of MAPK, as was the induction of cyclin D1 (data not shown), consistent with previous results from our lab that have consistently found an obligatory role for MAPK signaling in the proliferative response to activated Ras and growth factors in human endothelial cells [10,21,22], as well as those previously reported by Munchhof et al. [8]. The effects of PI-3'-kinase inhibition were difficult to interpret, as inhibition of this signal also partially inhibited mTORC1 activation, likely explaining the intermediate and variable effect we observed with this inhibitor in proliferation assays (data not shown). However we found that at low doses of rapamycin (0.1 ng/ml), the NF1 mediated activation of AKT and ERK were unaffected while the NF1 mediated S6-phosphorylation was completely inhibited (Figs. 6A and S2). This dose of rapamycin was also sufficient to completely abrogate the proliferative response observed with the loss of NF1 expression. These data suggest an unexpected obligatory contribution of this pathway to the enhanced proliferation following suppression of NF1 expression. We next investigated the role mTORC1 in regulating two known modulators of endothelial cell cycle progression, Cyclin D1 and the phosphorylation of the corresponding cyclin dependent kinase inhibitor, p27^{KIP} [23]. We find the induction of cyclin D and the phosphorylation of p27 is strongly affected by low dose rapamycin. Collectively these data suggest that the proliferative response we observe in endothelial cells with reduced NF1 expression is highly sensitive to inhibition by rapamycin which seems to act at least in part by suppressing cyclin D induction and the phosphorylation of p27.

As cells lacking NF1 induce a hyperplastic phenotype in co-culture, we reasoned that this abnormal morphogenesis might be altered by low dose rapamycin. To test this, control and NF1-kd cells were co-cultured with primary fibroblasts, with or without low doses of rapamycin (Fig. 7). We found that in control endothelial cells the presence of rapamycin inhibited the number of tube-like structures; however the general characteristics of these branching networks were similar to untreated cells. Interestingly, in the NF1-kd cells, the sheet like phenotype was not present, rather the cells underwent normal branching morphogenesis, indicating that the blunting of mTORC1 signaling and endothelial cell proliferation was able to restore a normal phenotype to these cells.

Discussion

Our data strongly support a critical role for NF1 in the regulation of the vascular endothelium. These findings are in agreement with previous studies done in developmental models whereby the endothelial specific deletion of NF1 resulted in embryonic lethality. These data also agree with the data of Munchhof et al. [8], who found that knocking down of expression in endothelial cells augmented VEGF and FGF signaling and induced angiogenesis. Importantly our data extend this previous work in several critical areas both conceptually and mechanistically.

Notably, our data demonstrate that the loss of NF1 is sufficient to induce activation of Ras and initiate downstream signaling. This suggests that NF1 doesn't just play a passive role in dampening Ras activation but rather is an active regulator of GTP-Ras accumulation. The loss of NF1 triggers the accumulation of Ras-GTP even under serum and growth factor-free conditions. The resulting accumulation of Ras-GTP is sufficient to initiate changes in cell behavior, including entry into the cell cycle and enhanced growth rates under mitogenic conditions. Importantly, we find no evidence of the senescence associated with the loss of NF1 in other cell types [11]. This finding is consistent with our previous findings showing a lack of senescence following expression of activated H-Ras. The changes in Ras-related signaling result in a gross perturbation of the endothelial cell vasculogenic program. Under co-culture conditions, normal primary endothelial cells stop growing, elongate, and form interconnecting tubules; NF1-kd cells however, continue to maintain at least a partially proliferative phenotype and fail to branch and form tubular structures reliably. To our knowledge this is the first report that the loss of NF1 is sufficient to alter the morphogenic program of endothelial cells. This finding may help to shed light on the underlying cause of at least some of the vascular anomalies seen in some NF1 patients and may provide an *in vitro* model to study the molecular basis for these vascular defects.

It is noteworthy that the alteration of the morphogenic phenotype appears to be quite plastic. Under conditions where the vessel structures are malformed, a return of normal Ras regulation permits at least a partial normalization. Similarly even in stable vessel structures, the loss of NF1 and accompanying Ras activation results in the emergence of an altered phenotype. These data suggest that in NF1 patients, the acquisition of an additional mutation or epigenetic silencing of the remaining copy of NF1 might be sufficient to trigger Ras activation, autonomous proliferation and abnormal vessel formation – even in post-developmental quiescent vasculature. This data begins to explain the high frequency of moyamoya disease, arteriovenous malformations and possibly even aneurysms in the NF1 patient population. The finding that the Ras-GAP, RASA1 (p120-GAP), is mutated in several distinct vascular anomalies [24,25] including capillary malformation-arterial venous malformations and Vein of Galen aneurysms supports this hypothesis, though it is currently unclear if the loss of RASA1 is sufficient to result in Ras activation. Recent data investigating the role of RASA1 in the endothelium of adult mice suggests that loss of RASA1 does result in abnormal morphology and lymphatic endothelial proliferation. However the activation of Ras and Ras-related signaling, along with the abnormal vascular phenotype were dependent upon coincident VEGFR3 signaling [26]. Current experiments are determining if the homozygous loss of NF1 in the post-natal vasculature is sufficient to trigger Ras signaling and vascular anomalies, particularly in a haploinsufficient microenvironment which is known to be pro-angiogenic [7].

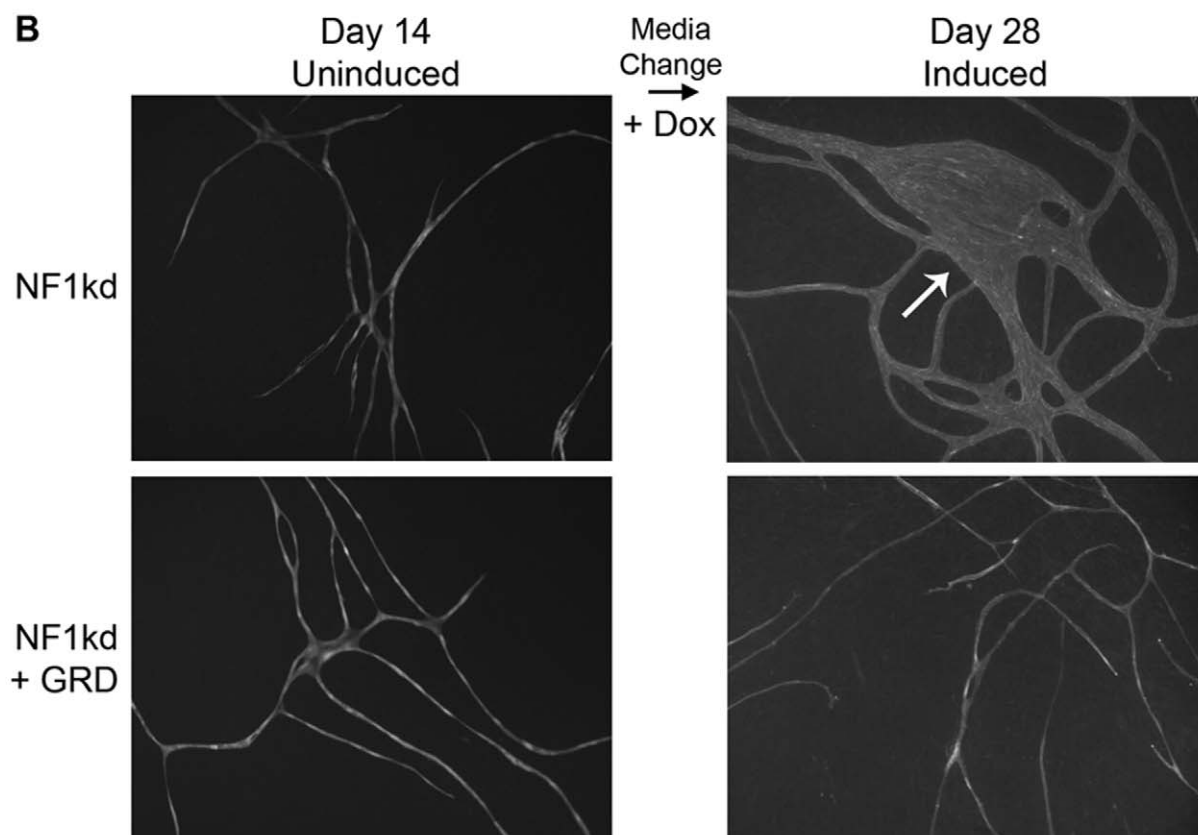
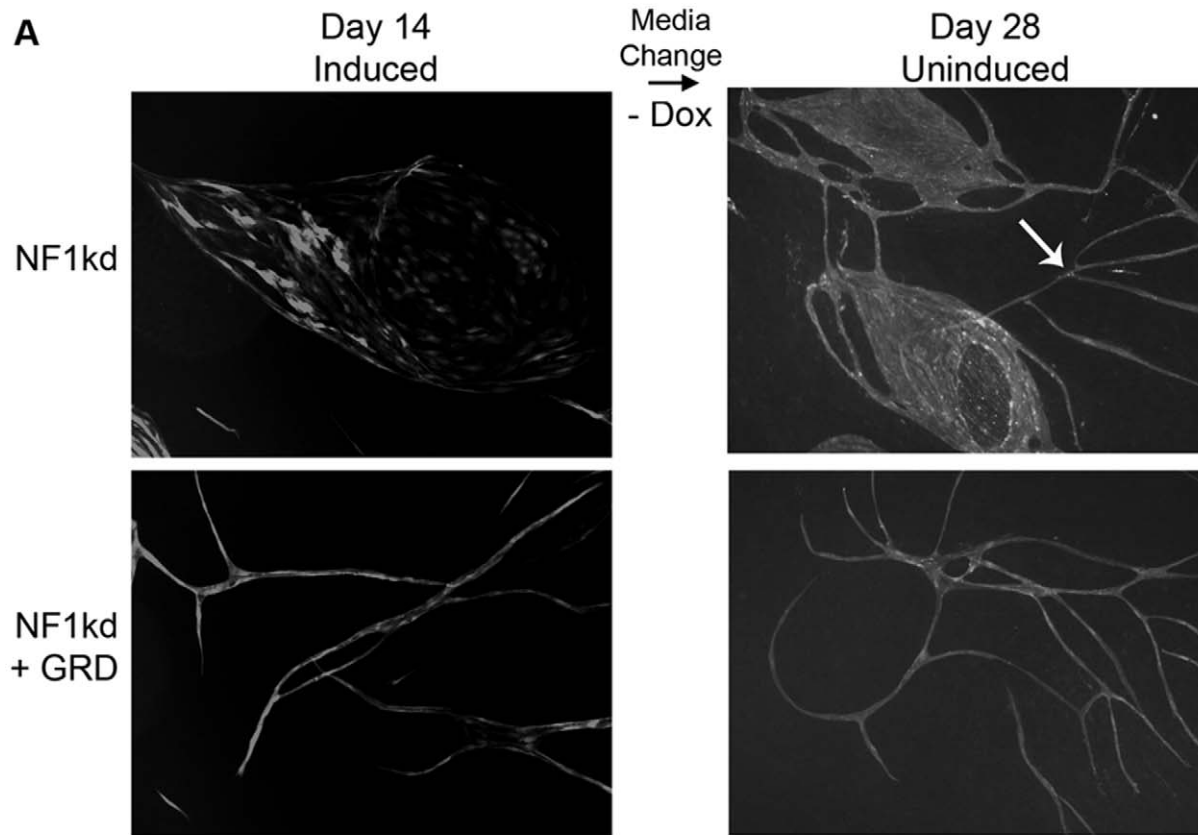


Figure 5. Changes in vascular morphogenesis are reversible and dependent upon active Ras. Cells for these experiments were co-infected with TRIPZ-NF1KD and pSLIK virus that was either empty or expressing the GRD domain of NF1 and sorted by FACS for double positive populations. These cells were then plated in co-culture with primary fibroblasts in the presence or absence of doxycycline as indicated. (A) Admixed cultures were allowed to form structures for 14 days. Representative fields were photographed using expressed RFP at this time and doxycycline was removed from the culture medium. Co-cultures were allowed to persist for an additional 14 days, after which time representative fields were again photographed following visualization of endothelial cells with FITC-labeled UEA-1 lectin. (B) Double positive cells were plated in the absence of doxycycline for 14 days and endothelial cells were visualized by staining live cultures with FITC labeled UEA-1 lectin and representative fields were photographed. Doxycycline was then added to cultures and they were incubated for an additional 14 days followed by visualization again using UEA-1 lectin. Arrows in both (A) and (B) highlight representative changes in morphology.
doi:10.1371/journal.pone.0049222.g005

Our data directly addressed the role of Ras in the endothelial cell regulation by NF1. While some cell types have been reported to have important Ras-independent functions of NF1 [27,28,29], our data suggest that alteration in Ras signaling is essential for the effects seen upon loss of NF1 in endothelial cells. We found that all of the phenotypes and signaling changes observed following the loss of NF1 were rescued by re-expression of the GAP-related domain of NF1 and the restoration of Ras regulation. However it is currently unclear if there is isoform specificity to the Ras activation seen following the loss of NF1 or if particular isoforms are linked to the phenotypic manifestations. Moreover we cannot rule out that other signaling changes are also taking place and contributing to the observed phenotypes.

We find that loss of NF1 triggered several Ras-related signaling pathways including activation of PI-3'-kinase and activation of mTOR signaling. The activation of mTOR was obligatory for the autonomous proliferation triggered by the loss of NF1. The proliferative dependency appears to arise out of mTOR-dependent phosphorylation of p27 leading to its degradation as well as induction of cyclin D1 which has previously been implicated in NF1 tumors [30]. Previous studies in our lab have determined that p27 degradation is an essential step in endothelial progression to S-Phase [23]. Our data do not address whether mTORC1 is directly phosphorylating p27, however we did not find any significant inhibition of either AKT or ERK (two enzymes known to phosphorylate p27) by the low dose of rapamycin we employed. It

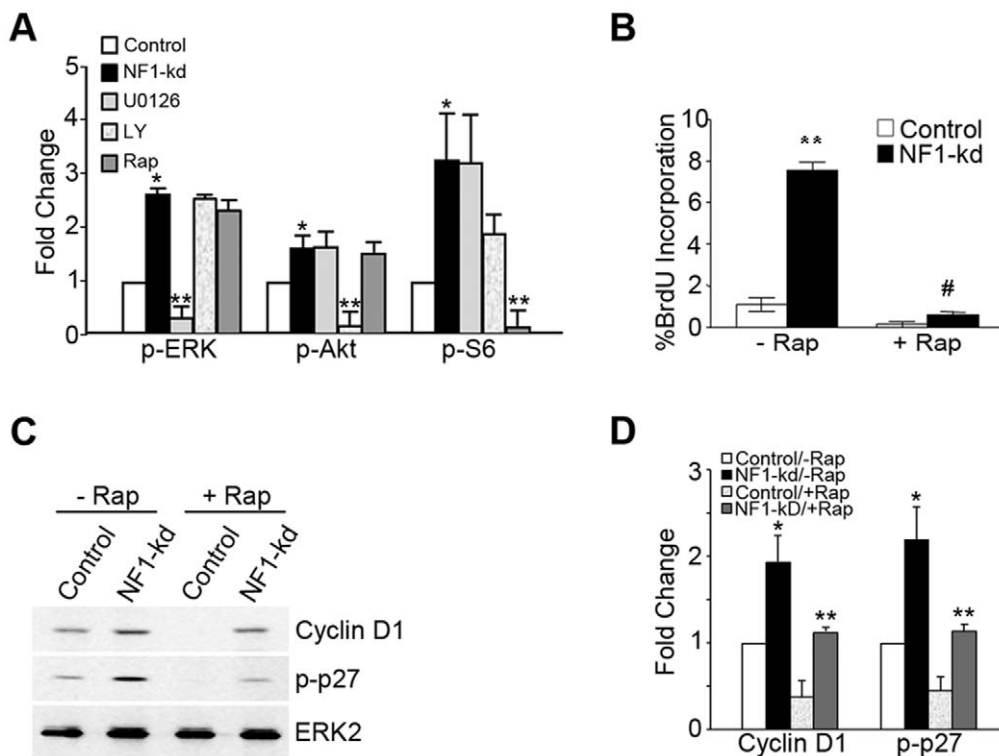


Figure 6. mTORC1 is critical for enhanced proliferation following the loss of NF1 (pSIREN). Control and NF1 knockdown primary endothelial cells were serum deprived for 24 h prior. Inhibitors were added at the time of serum starvation at the following concentrations (U0126, 1 μ M; LY29002, 100 nM; Rapamycin, 0.1 ng/ml). At 21 h, some cells were pulsed with BrdU for 3 h. At 24 h cell lysates were made and probed for changes in cellular signaling (A) using phospho-specific antibodies or changes in cellular proliferation (B), visualized with an anti-BrdU antibody. (A) Results from three independent experiments were quantified from western blots similar to those shown in Supplemental 2. Data represent band average response under each condition. Error bars represent standard error of the mean and relevant statistical relationships are shown (* p <0.05 compared to Control; ** p <0.01 compared to NF1-kd). (B) BrdU positive cells were quantified and the data was graphed as the % positive cells compared to total cell number. The error bars represent standard error of the mean (** p <0.01 compared to control; # p <0.01 compared to NF1-kd). In a similar experiment, cells were serum starved 24 h in the absence and presence of rapamycin (0.1 ng/ml) and lysates were made and probed with antibodies against cyclin D1, phospho-p27 and ERK2, the latter used as a loading control. (D) Results from three experiments similar to those performed in (C) were quantified. Data represents the mean result. Error bars represent standard error of the mean (* p <0.05 compared to control; ** p <0.01 compared to NF1-kd).
doi:10.1371/journal.pone.0049222.g006

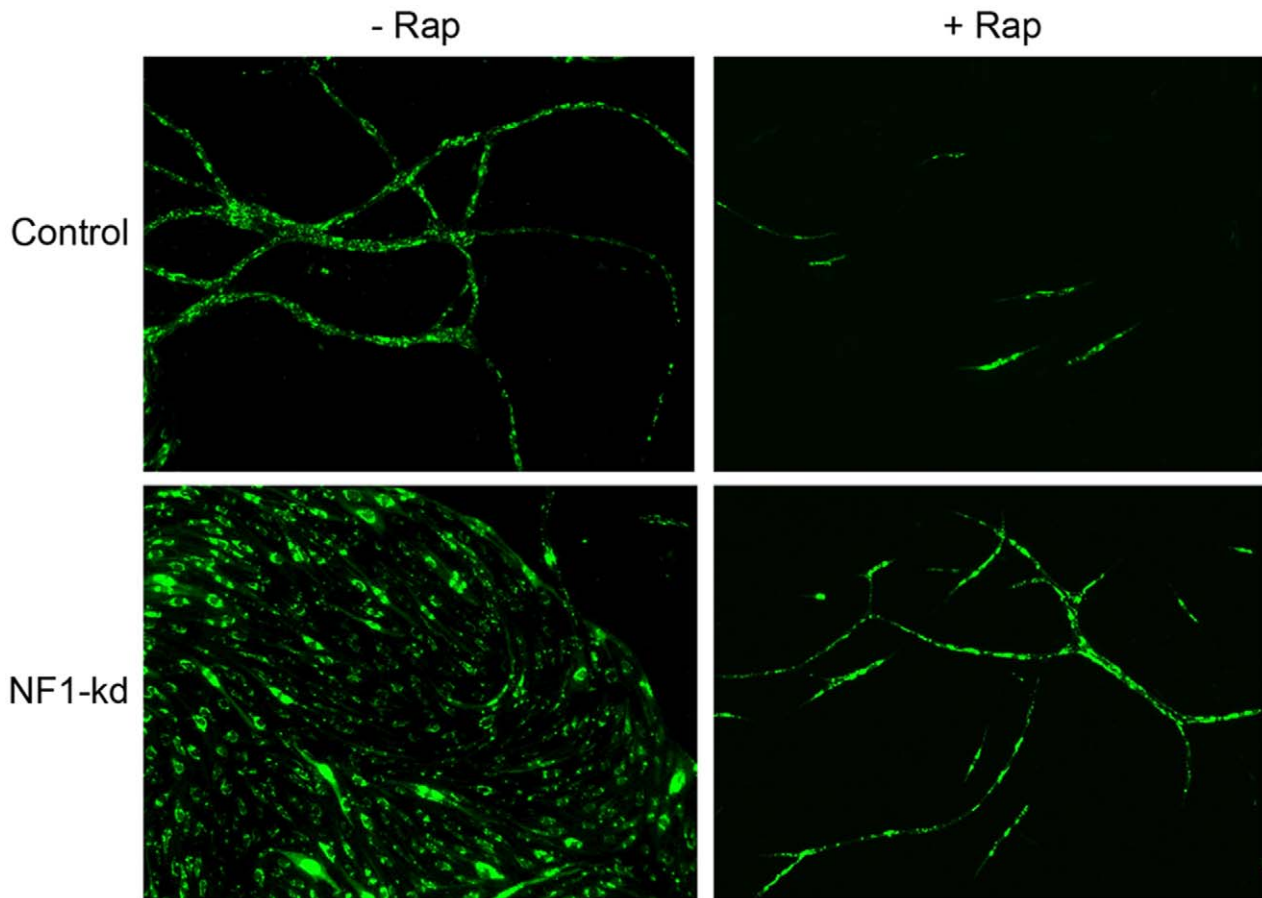


Figure 7. Abnormal vascular morphogenesis is normalized by Rapamycin. Endothelial cells infected with pSIREN expressing either non-targeting (control) or shRNA directed against NF1 (NF1-kd) were plated in co-culture with primary fibroblasts. Co-cultures were incubated in the presence of vehicle (DMSO) or Rapamycin (0.1 ng/ml) from the time of plating. Vascular structures are shown at day 14, visualized by expression of GFP in the endothelial cells by the pSIREN vector.
doi:10.1371/journal.pone.0049222.g007

is interesting to note that activation of mTOR has been associated with cutaneous vascular malformations [31]. The sensitivity of the NF1-kd endothelial cell proliferation and the normalization of the vascular morphogenesis following treatment with low-dose rapamycin in our assays suggest that “Rapalogs” currently in clinical trials as anti-tumor medications may be an effective management tool for certain types of vascular dysfunction in NF1 patients. In addition, it seems likely that these patients may be well-positioned to benefit from other currently approved vasculoprotective therapeutics, e.g., statins which can dampen both Ras and mTOR signaling [32,33] as well as metformin which activates AMPK signaling [34] to dampen mTOR.

Materials and Methods

Ethics Statement

Parents and legal guardians of donors provided consent that discarded tissue could be used for research purposes. The collection and use of tissue samples was evaluated and approved by the Institutional Review Board of Albany Medical Center. It was not considered Human subjects research under 45 CFR (Basic HHS Policy for Protection of Human Research Subjects) part 46 because, (1) the specimens were not collected specifically for the currently proposed research project and (2) the investigator(s) cannot readily ascertain the identity of the individual(s), consistent

with the guidelines set forth by the Office of Human Research Protection of the US Federal Government.

Cell Culture

HUVECs (Human Umbilical Vein Endothelial Cells) were purchased from Cascade Biologics (Portland, Oregon, USA) or Lifeline Cell Technologies (Fredrick MD, USA). Cells were cultured as previously described [21]. Cells were made quiescent by incubation in serum free MCDB-131 supplemented with 1% penicillin/streptomycin and 2 mM L-glutamine (SF), where noted. Stimulation was performed with complete growth media. Primary fibroblasts were isolated from human foreskins provided as de-identified, discarded tissue from neonatal circumcision procedures at Albany Medical Center and grown in DMEM containing 10% FBS and 1% penicillin/streptomycin.

Western Blotting

Western blotting analysis used the following antibodies: mouse anti-pERK (Santa Cruz Biotechnology), rabbit anti-ERK2 (Santa Cruz Biotechnology), rabbit anti-NF1 (Bethyl Laboratories), rabbit anti-p27 (Santa Cruz Biotechnology), rabbit anti-phospho-p27 (Zymed), rabbit anti-pS6, rabbit anti-pAKT, mouse anti-cyclin D1, mouse anti-pan Ras (Oncogene Research, Calbiochem). All antibodies were used at a dilution of 1:1000 overnight at 4°C. Other conditions were the same as described in [21] except

exposures were captured on a Kodak 4000 MM imager. All figures are representative of at least three independent experiments. Quantification was performed by using volume measurements from the KODAK imager for the band of interest and normalizing to the volume of the loading control (ERK2) for that lane. Data is the mean of three experiments and error bars represent the standard error of the mean. Statistical analysis was performed using students t-test or in some cases ANNOVA with a Bonferroni correction for multivariable conditions.

Plasmid Construction

To stably knockdown *NF1* expression in HUVECs a shRNA sequence targeting *NF1* (5'gatccGGACACAATGAGATTA-GATTTCTCAAGAGAAAATCTAATCT-CATTGTGTCTTTTTTACGCGTg3' sense strand) and (5'aattcACGCGTAAAAAAGGACACAATGAGATTA-GATTTTCTCTTGAGAAATCTAATCTCATTGTGTCTCCg3' antisense strand) into the RNAi-Ready pSIREN-RetroQ-ZsGreen retroviral expression vector from Clontech. The shRNA sequences were synthesized by Operon and had a MluI restriction site in the hairpin loop region with BamHI and EcoRI cut overhangs for cloning. The oligonucleotides were annealed and ligated into the BamHI/EcoRI cut pSIREN-ZsGreen according to the "Knockout RNAi Systems User Manual" from Clontech. The same annealed shRNAs were ligated into the BamHI/EcoRI cut pSIREN-RetroQ-DsRed-Express vector to make a knockdown vector expressing red fluorescent protein. A Negative Control shRNA annealed oligonucleotide provided by Clontech was ligated similarly into the pSIREN-RetroQ-ZsGreen to make a control vector. Inducible knockdown of NF1 was achieved using a lentiviral pTRIPZ vector from Open Biosystems carrying the V2THS_260806 sequence for knocking down NF1. Schematics of these vector constructs are shown in *Supplementary Fig. S1*. To make a lentiviral vector expressing the gap related domain (GRD) of NF1 the MSCV-puro-GRD-V5 plasmid was purchased from Addgene. A *BglIII/NotI* fragment carrying the GRD was ligated into *BamHI/NotI* cut pEN_TRE2 [10]. The Tet promoter along with NF1-GRD was then put into the Gateway compatible destination vector pSLIK-Venus [35] using an LR-ClonaseTM reaction resulting in the pSLIK-GRD-Venus plasmid.

Production and Infection with Retroviruses and Lentiviruses

The pSIREN-RetroQ-ZsGreen lentiviral vector carrying a negative control or a NF1 shRNA was transfected into retroviral packaging PhoenixA cells [36] using Lipofectamine 2000 reagent. The media from the transfected cells was sterile filtered through a 0.4 μ M filter and the viral supernatant was used to infect low passage HUVECs in the presence of 5 μ g/mL of polybrene. The HUVECs underwent a second round of infection in a similar manner after 24 h. The infection efficiency was 70–80% and a pure population of infected cells was obtained by flow cytometry based sorting under sterile conditions, using ZsGreen as a selectable marker. The lentiviral vectors (2 μ g) were co-transfected along with the respective packaging plasmids into 60% confluent 293 FT packaging cells (Invitrogen) cell using Lipofectamine 2000 reagent (Invitrogen). The pSLIK based vectors were co-transfected with three 3rd generation packaging plasmids, 3 μ g each of pMDLg/pRRE(Addgene, plasmid 12251), pRSVREV (Addgene, plasmid 12253), the vesicular stomatitis virus (VSV) G envelope plasmid pVSV (Addgene, plasmid 12259) [37]. The pTRIPZ lentiviral vectors were co-transfected along with two 2nd genera-

tion packaging plasmids, 3 μ g each of pCMV-dR8.2 dvpr (Addgene, plasmid 8455), pCMV-VSVG (Addgene, plasmid 8454). Low passage HUVECs were infected and sorted in a similar manner as above to obtain a pure population of knockdown cells. With both the retroviral and lentiviral vectors, three independent infections were performed on three independent endothelial cell cultures in order to insure representative results.

Measurement of DNA Synthesis and Growth Assays

Endothelial cells were serum starved for 24 h, after which complete growth media (GM) was added as a mitogenic stimulus for 16 h. Measurements of BrdU incorporation were performed as previously described [22]. In some cases indicated doses of Rapamycin were added to the cells at the time of serum starvation. Growth assays were conducted as described previously [38]. Population doublings were calculated using the formula: Population Doublings = Log(Final cell number/Initial cell number)/Log2. Cumulative population doublings represent the sum of population doublings from all previous passages.

Co-culture Assay

This assay was performed as previously described [15] with modifications as we have previously reported [10]. Cell were typically tracked using the expression of fluorescent markers introduced during genetic modification. In some cases, cells were stained live with a FITC-tagged UEA-1 lectin (Sigma-Aldrich) or fixed in 3.7% formaldehyde and visualized with UEA-1 lectin.

Supporting Information

Figure S1 Schematic representation of NF1 knockdown constructs. (TIF)

Figure S2 Knockdown of NF1 by pSiren activates Ras and cellular signaling. (A) Cells infected with the pSiren vector targeting a control sequence or NF1 (NF1-kd) were analyzed by western blotting for the expression of NF1 using ERK2 as a loading control (*upper panels*). Cell lysates were also probed for active Ras using GTP pull-down experiments using anti-Ras immunoblots (*lower panels*). GTP-Ras represents Ras bound to GST-Raf beads. Levels of total Ras in the input lysate were used to insure similar levels of lysate loading onto the beads. (B) Cell lysates from pSiren infected cells expressing control or NF-kd shRNAs were analyzed for presence of signals known to be downstream of active Ras and for sensitivity to treatment with low doses of the signal transduction inhibitors indicated using immunoblotting. ERK2 is used as a loading control to insure equivalent cellular lysate. Quantification of several similar experiments is shown in Figure 6A. (TIF)

Acknowledgments

The authors would like to thank Dr. Peter Vincent and Dr. Harold Singer for critical reading of the manuscript.

Author Contributions

Conceived and designed the experiments: AB Q-FL KP. Performed the experiments: AB Q-FL QZ. Analyzed the data: AB Q-FL QZ KP. Contributed reagents/materials/analysis tools: AB Q-FL. Wrote the paper: AB Q-FL KP.

References

- McClatchey AI (2007) Neurofibromatosis. *Annu Rev Pathol* 2: 191–216.
- Friedman JM, Arbiser J, Epstein JA, Gutmann DH, Huot SJ, et al. (2002) Cardiovascular disease in neurofibromatosis 1: report of the NF1 Cardiovascular Task Force. *Genet Med* 4: 105–111.
- Rasmussen SA, Yang Q, Friedman JM (2001) Mortality in neurofibromatosis 1: an analysis using U.S. death certificates. *Am J Hum Genet* 68: 1110–1118.
- Xu J, Ismat FA, Wang T, Yang J, Epstein JA (2007) NF1 regulates a Ras-dependent vascular smooth muscle proliferative injury response. *Circulation* 116: 2148–2156.
- Lasater EA, Li F, Bessler WK, Estes ML, Vemula S, et al. (2010) Genetic and cellular evidence of vascular inflammation in neurofibromin-deficient mice and humans. *J Clin Invest* 120: 859–870.
- Gitler AD, Zhu Y, Ismat FA, Lu MM, Yamauchi Y, et al. (2003) Nf1 has an essential role in endothelial cells. *Nat Genet* 33: 75–79.
- Wu M, Wallace MR, Muir D (2006) Nf1 haploinsufficiency augments angiogenesis. *Oncogene* 25: 2297–2303.
- Munchhof AM, Li F, White HA, Mead LE, Krier TR, et al. (2006) Neurofibroma-associated growth factors activate a distinct signaling network to alter the function of neurofibromin-deficient endothelial cells. *Hum Mol Genet* 15: 1858–1869.
- Dasgupta B, Dugan LL, Gutmann DH (2003) The neurofibromatosis 1 gene product neurofibromin regulates pituitary adenylate cyclase-activating polypeptide-mediated signaling in astrocytes. *J Neurosci* 23: 8949–8954.
- Bajaj A, Zheng QX, Adam A, Vincent P, Pumiglia K (2010) Activation of endothelial Ras signaling bypasses senescence and causes abnormal vascular morphogenesis. *Cancer Res* 70: 3803–3812.
- Courtois-Cox S, Genter Williams SM, Reczek EE, Johnson BW, McGillicuddy LT, et al. (2006) A negative feedback signaling network underlies oncogene-induced senescence. *Cancer Cell* 10: 459–472.
- Dasgupta B, Yi Y, Chen DY, Weber JD, Gutmann DH (2005) Proteomic analysis reveals hyperactivation of the mammalian target of rapamycin pathway in neurofibromatosis 1-associated human and mouse brain tumors. *Cancer Res* 65: 2755–2760.
- Johannessen CM (2005) The NF1 tumor suppressor critically regulates TSC2 and mTOR. *PROC NATL ACAD SCI* 102: 8573–8578.
- Johannessen CM, Johnson BW, Williams SM, Chan AW, Reczek EE, et al. (2008) TORC1 is essential for NF1-associated malignancies. *Curr Biol* 18: 56–62.
- Bishop ET, Bell GT, Bloor S, Broom IJ, Hendry NF, et al. (1999) An in vitro model of angiogenesis: basic features. *Angiogenesis* 3: 335–344.
- Mavria G, Vercoulen Y, Yeo M, Paterson H, Karasarides M, et al. (2006) ERK-MAPK signaling opposes Rho-kinase to promote endothelial cell survival and sprouting during angiogenesis. *Cancer Cell* 9: 33–44.
- Hayward SW, Wang YH, Cao M, Hom YK, Zhang BH, et al. (2001) Malignant transformation in a nontumorigenic human prostatic epithelial cell line. *Cancer Res* 61: 8135–8142.
- Banerjee S, Crouse NR, Emmett RJ, Gianino SM, Gutmann DH (2011) Neurofibromatosis-1 regulates mTOR-mediated astrocyte growth and glioma formation in a TSC/Rheb-independent manner. *Proc Natl Acad Sci USA* 108: 15996–16001.
- Lee da Y, Yeh TH, Emmett RJ, White CR, Gutmann DH (2010) Neurofibromatosis-1 regulates neuroglial progenitor proliferation and glial differentiation in a brain region-specific manner. *Genes Develop* 24: 2317–2329.
- Ma J, Li M, Hock J, Yu X (2012) Hyperactivation of mTOR critically regulates abnormal osteoclastogenesis in neurofibromatosis Type 1. *J Orthopaedic Res* 30: 144–152.
- Meadows KN, Bryant P, Pumiglia K (2001) Vascular endothelial growth factor induction of the angiogenic phenotype requires Ras activation. *J Biol Chem* 276: 49289–49298.
- Meadows KN, Bryant P, Vincent PA, Pumiglia KM (2004) Activated Ras induces a proangiogenic phenotype in primary endothelial cells. *Oncogene* 23: 192–200.
- Bryant P, Zheng Q, Pumiglia K (2006) Focal adhesion kinase controls cellular levels of p27/Kip1 and p21/Cip1 through Skp2-dependent and -independent mechanisms. *Mol Cell Biol* 26: 4201–4213.
- Eerola I, Boon LM, Mulliken JB, Burrows PE, Domp Martin A, et al. (2003) Capillary malformation-arteriovenous malformation, a new clinical and genetic disorder caused by RASA1 mutations. *Am J Hum Genet* 73: 1240–1249.
- Revenu N, Boon LM, Mulliken JB, Eniolas O, Cordisco MR, et al. (2008) Parkes Weber syndrome, vein of Galen aneurysmal malformation, and other fast-flow vascular anomalies are caused by RASA1 mutations. *Human Mutation* 29: 959–965.
- Lapinski PE, Kwon S, Lubeck BA, Wilkinson JE, Srinivasan RS, et al. (2012) RASA1 maintains the lymphatic vasculature in a quiescent functional state in mice. *J Clin Invest* 122: 733–747.
- Brown JA, Diggs-Andrews KA, Gianino SM, Gutmann DH (2012) Neurofibromatosis-1 heterozygosity impairs CNS neuronal morphology in a cAMP/PKA/ROCK-dependent manner. *Mol Cell Neurosci* 49: 13–22.
- Brown JA, Gianino SM, Gutmann DH (2010) Defective cAMP generation underlies the sensitivity of CNS neurons to neurofibromatosis-1 heterozygosity. *J Neurosci* 30: 5579–5589.
- Ismat FA, Xu J, Lu MM, Epstein JA (2006) The neurofibromin GAP-related domain rescues endothelial but not neural crest development in Nf1^{-/-} mice. *J Clin Invest* 116: 2378–2384.
- Johannessen CM, Johnson BW, Williams SM, Chan AW, Reczek EE, et al. (2008) TORC1 is essential for NF1-associated malignancies. *Curr Biol* 18: 56–62.
- Shirazi F, Cohen C, Fried L, Arbiser JL (2007) Mammalian target of Rapamycin (mTOR) is activated in cutaneous vascular malformations in vivo. *Lymphat Res Biol* 5: 233–236.
- Endres M, Laufs U (2004) Effects of statins on endothelium and signaling mechanisms. *Stroke* 35: 2708–2711.
- Prinz V, Endres M (2011) Statins and stroke: prevention and beyond. *Curr Opin Neurol* 24: 75–80.
- Ewart MA, Kennedy S (2011) AMPK and vasculoprotection. *Pharmacol Therap* 131: 242–253.
- Shin KJ, Wall EA, Zavzavadjian JR, Santat LA, Liu J, et al. (2006) A single lentiviral vector platform for microRNA-based conditional RNA interference and coordinated transgene expression. *Proc Natl Acad Sci USA* 103: 13759–13764.
- Kinsella TM, Nolan GP (1996) Episomal vectors rapidly and stably produce high-titer recombinant retrovirus. *Human Gene Ther* 7: 1405–1413.
- Dull T, Zufferey R, Kelly M, Mandel RJ, Nguyen M, et al. (1998) A third-generation lentivirus vector with a conditional packaging system. *J Virol* 72: 8463–8471.
- Serrano M, Lin AW, McCurrach ME, Beach D, Lowe SW (1997) Oncogenic ras provokes premature cell senescence associated with accumulation of p53 and p16INK4a. *Cell* 88: 593–602.